



Implementation of practices shapes the effectiveness of agricultural diversification for arthropod related ecosystem services: a meta-analysis

Gaëtan Seimandi-Corda^{1,2} · Chloe MacLaren^{1,3,4} · Kevin Tougeron⁵ · Johannes Forkman⁴ · Jess Hood¹ · Andrew Mead¹ · Amelia Dixon¹ · Samantha M. Cook¹

Received: 12 May 2025 / Accepted: 10 December 2025
© The Author(s) 2026

Abstract

Agricultural intensification has increased food production but has also caused significant environmental degradation and biodiversity loss. Diversification practices can mitigate these impacts while sustaining yields. While previous meta-analyses have examined their effects on arthropod populations and associated ecosystem services, focusing on individual practices or treating them as a whole, this is the first study to compare a wide range of diversification strategies across specific arthropod groups and assess how management factors, such as plant diversity, spatial configuration, and sowing time, modulate their outcome. Clarifying these mechanisms is crucial for optimizing diversification practices and enhancing their adoption by farmers. We conducted the most up-to-date and comprehensive meta-analysis to evaluate the effects of four diversification practices (agroforestry, intercropping, flower resource addition, and maintaining semi-natural habitats) on arthropods (parasitoids, predators, pollinators, and herbivores), their associated ecosystem services (predation, parasitism, pollination), and disservices (herbivory), using 19,421 data points from 449 publications. On average, all diversification practices increased beneficial arthropod populations and their services by 41%, compared to a non-diverse control, and reduced herbivore abundance and plant damage by 33%. Intercropping was particularly effective, reducing herbivore abundance and damage by 39% and 30%, respectively, while increasing predator and parasitoid populations by 48% and 56%. Other practices showed no consistent effects, likely due to high variability across studies. For the first time, we tested the influence of management factors and found complex effects on parasitoid abundance and parasitism, highlighting the need for context-specific approaches. These results highlight the importance of tailored policies and targeted research to support the adoption of diversification practices. Effective implementation can reduce reliance on insecticides while simultaneously promoting ecosystem services and supporting the transition to sustainable agricultural systems.

Keywords Plant diversity · Pests · Flower margin · Intercropping · Insect

1 Introduction

Agricultural intensification, achieved through increasing mechanization, high inputs of fertilizers and pesticides, expanding field sizes, and reducing the number of crops grown, has resulted in a remarkable surge in food production (Grassini et al. 2013). However, agricultural intensification is also recognized as a primary driver of global environmental changes, exerting substantial adverse impacts on biodiversity (Raven and Wagner 2021). These biodiversity impacts, in turn, affect the provision of essential ecosystem services required to support agricultural production, including pest regulation and crop pollination (Ratnadass et al. 2012; Emmerson et al. 2016; Kass 2020), thereby posing a global threat to food production. Consequently, it is

✉ Gaëtan Seimandi-Corda
gaetan.seimandi@outlook.fr

¹ Rothamsted Research, Harpenden, Hertfordshire, UK

² UMR AGIR, Université de Toulouse, INRAE, Castanet-Tolosan F-31326, France

³ International Maize & Wheat Improvement Centre (CIMMYT), Southern Africa Regional Office (SARO), P.O. Box MP163, Harare, Zimbabwe

⁴ Department of Crop Production Ecology, Swedish University of Agricultural Sciences, Uppsala 75007, Sweden

⁵ Ecology of Interactions and Global Change Laboratory, Institute for Biosciences, Université de Mons, Mons, Belgium

imperative to develop sustainable farming practices that can sustain yields while minimizing impact on the environment.

In recent years, agricultural diversification, especially practices where multiple plant species are grown simultaneously, has been advocated to reduce the negative impact of farming practices on biodiversity while maintaining high productivity (Kremen et al. 2012; Isbell et al. 2017; Brooker et al. 2023; Bommarco 2024). Diversification can encompass an increase in crop diversity temporally through longer and more varied crop rotations, as well as spatially via intercropping, or an increase in non-crop plant diversity through the addition of flower margins, agroforestry, or the maintenance of semi-natural habitats (SNH), such as hedgerows proximate to crops (Kremen et al. 2012; Hufnagel et al. 2020; Wan et al. 2020). Global syntheses have shown that plant diversification practices increase biodiversity compared to highly simplified cropping systems (Lichtenberg et al. 2017; Dainese et al. 2019), and also that they can have positive effects on water and soil quality, while reducing greenhouse gas emissions (Tamburini et al. 2020; Beillouin et al. 2021). Moreover, diversification practices have exhibited beneficial impacts on ecosystem services essential to agriculture, including pest and disease management, and the promotion of insect pollinators (Wan et al. 2020; Zamorano et al. 2020).

Ecosystem services related to the populations and activities of insects and other arthropods are critical to support agricultural production. On one hand, insect pests cause approximately 18% of global yield losses (Oerke 2006), while on the other hand, insect pollinators contribute to around 8% of food production (Aizen et al. 2009). Crop diversification can affect arthropods via both top-down and bottom-up effects. Bottom-up effects occur when plant diversity has a direct effect on herbivores, altering their populations or the amount of damage they cause to crops. Such effects can be observed when additional plant species disturb host plant location, visually hide the crop, or have a repellent or attractive effect on herbivores (Ratnadass et al. 2012). Top-down effects occur when plant diversity supports biocontrol agents (i.e., predators and parasitoids) by providing resources such as pollen, nectar, alternative hosts and prey, or habitat and refugia (Ratnadass et al. 2012). These resources support the population of beneficial insects that suppress herbivore numbers, thereby ultimately boosting crop yield or reducing the need for pesticides to sustain yields. The addition of plant species providing floral resources will also support pollinator populations and enhance the yield of some crops (Nicholls and Altieri 2013).

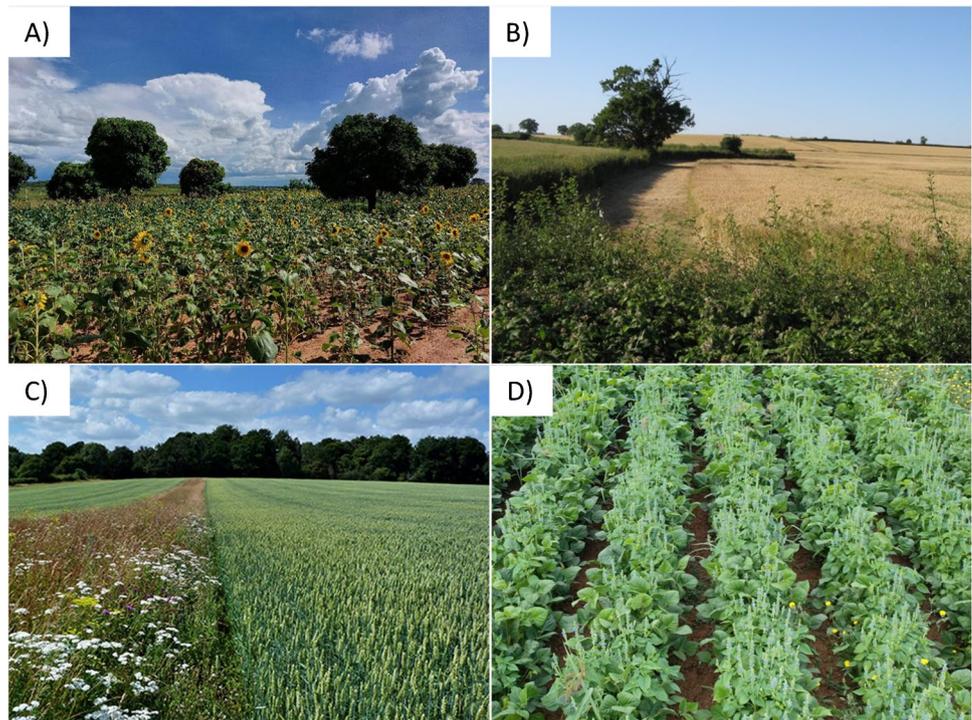
Previous syntheses that considered all plant diversification practices together have shown a trend of positive effects on beneficial insects (predators, parasitoids, and pollinators) and their associated services, as well as a decrease in herbivore abundance and crop damage (Lichtenberg et al. 2017; Wan et al. 2020, 2022; Tamburini et al. 2020). Some

syntheses have focused on specific practices such as intercropping (Iverson et al. 2014; Rakotomalala et al. 2023), agroforestry (Pumariño et al. 2015; Perez-Alvarez et al. 2023), or the addition of flower resources or SNH (Albrecht et al. 2020; Crowther et al. 2023; Jachowicz and Sigsgaard 2025) and have generally reached similar conclusions. These studies give a generally positive view of diversification practices, yet their implementation by farmers is still limited. Reasons behind the barriers for adoptions of diversification practices are multiple and complex involving economic, sociological, and psychological factors but also limitations in the availability of specific knowledge about the effect of diversification practices (Blesh et al. 2023). Previous studies have not compared the effects of different diversification practices. A few have compared specific practices, for example, SNH with flower resource addition (Albrecht et al. 2020), or intercropping, agroforestry, and the presence of hedgerows (Beillouin et al. 2021; Jaworski et al. 2023). However, these studies focused on a narrow range of responses such as pest regulation and pollination services (Albrecht et al. 2020), or pest (including pathogen) regulation only (Beillouin et al. 2021; Jaworski et al. 2023). They did not distinguish between the abundance of the arthropods and the services they support, nor between effects on different functional groups such as parasitoids and predators. A comprehensive evaluation of the effects of diversification on the arthropod community as a whole, and the services and disservices provided, is therefore still lacking.

Moreover, the manner in which diversification practices are implemented and managed by farmers, including factors such as the number of added plant species and their spatial configuration, has often been overlooked. Only Albrecht et al. (2020) and Jachowicz and Sigsgaard (2025) examined the effect of the number of flower species. The first study also assessed how the age of the flower margin affected the services they studied. Rakotomalala et al. (2023) investigated the effect of different configurations of intercropping or combinations of plant families on pests and beneficials. The three studies showed that management factors are key moderators of the outcome of diversification practices. Understanding the effect of these management factors is crucial to provide specific knowledge to farmers and enable them to identify the most efficient practices and elucidate why certain farming systems yield better outcomes compared to others.

In this study, we focus on four farm- and landscape-level diversification practices: (1) intercropping, i.e., the practice of growing two or more crops in the same field at the same time; (2) agroforestry, where woody perennials and crops are grown on the same land; (3) the addition of flower resources, typically achieved through sown, non-woody flower strips or margins surrounding or within crops; (4) the presence of SNH, such as grasslands, woodlands, and hedgerows,

Fig. 1 Crop diversification practices considered in the analysis: **A** agroforestry (mango trees and sunflower), **B** maintenance of semi-natural habitats (hedgerow), **C** flower addition (flower strip along a cereal field), **D** intercropping (chia and mungo beans). Photocredit: Chloe MacLaren (A), Doug Lee (B), Sarah Hulmes (C), Fanny Raoux (D).



adjacent to the crop (Table S1). We reviewed articles published between 1980 and 2024 and collected 19,421 data points from 449 publications. Through meta-analysis, we aimed to take a closer look at differences in the effects of the four diversification practices on different groups of arthropods (herbivores, predators, parasitoids, and pollinators),

the services they support (predation, parasitism, and pollination), and their impact on the crop (crop damage via herbivory).

To address the critical knowledge gap regarding how best to implement diversification, we also tested the effect of key variables commonly used across studies to describe the

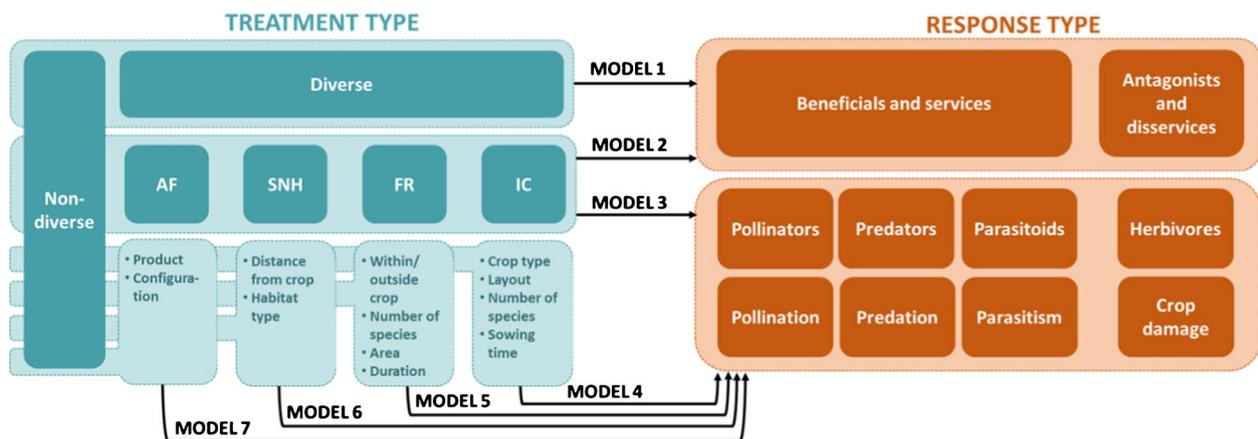


Fig. 2 Explanatory variables and levels used in models tested in a meta-analysis to examine the effect of different levels of plant diversification practices (agroforestry (AF), semi-natural habitat (SNH), flower resource addition (FR), intercropping (IC)) on beneficial (pollinators, predators, parasitoids) and antagonistic (herbivorous) arthropods and associated ecosystem services (pollination, predation, parasitism) or disservices (crop damage). For Models 1–3, light shaded boxes represent variables used in the models, with dark shaded boxes indicating the levels of these variables. Levels placed below each

other are nested with higher levels. “Non-diverse” was always the control level to which all other levels of plant diversification treatments were compared. For Models 4–7, the light shaded boxes for “treatment type” indicate each separate model, with text listing variables included in each model (levels not shown, see Table S1). Both the “treatment type” and “response type” variables are explanatory variables in each model, while the response variable always comprised the observations collected on arthropod abundances and service provision from the articles.

management of each diversification practice. These included (i) for intercropping: the number of intercropped species, the type of associated crops (cash crops or companions), sowing timings, and spatial configurations; (ii) for flower resources addition, the effect of flower species richness, patch size, duration, and proximity to the crop; (iii) for SNH, habitat type (hedgerows, woodland, grassland, grassy strips) and distance from the sampling location; (iv) for agroforestry systems, layout type (alley cropping and managed forest) and product (fruits and nuts, timber, no specific purpose).

2 Materials and methods

2.1 Data collection

Our search targeted articles presenting data collected in a context of agricultural plant diversification on the abundance of arthropods (herbivores, predators, parasitoids, pollinators), and/or on the provision of services (predation, parasitism, pollination) and disservices (herbivory) performed by arthropods. The diversification practices we considered were agroforestry, the addition of flower resources, intercropping, and the maintenance of SNH proximate to the crop (Fig. 1). The search was conducted using the Web of Science database (Core Collection), with the search string: (undersow* OR underseed* OR interseed* OR intercrop* OR “companion crop*” OR “companion plant*” OR “living mulch” OR interplant* OR “mixed crop” OR “flower strip*” OR “wildflower strip*” OR “flower margin” OR “field margin” OR “flower border” OR agroforestry OR “alley cropping” OR “trap crop*” OR hedgerow OR “field edge” OR “field boundaries” OR “crop diversification”) AND (insect OR pollinat* OR herbivor* OR predat* OR parasit* OR biocontrol OR “biological control” OR “pest control” OR “natural enem*”). Were removed from the search the terms: nematod* OR pathogen* OR slug*. It was undertaken for the first time on 7/01/20, and then updated regularly up to 11/06/24 to account for recently published articles (see Supporting information for details of the search) and accounted for studies published since 1980. In total, 3447 articles were identified using Web of Science.

Each paper was then carefully inspected, and data extracted if they reached the following six criteria: (1) The article deals with one of the diversification practices considered. (2) The article presents both high (treatment) and low (control) levels of plant diversity. Low diversity treatments were usually a monoculture (a single crop species grown on a plot) or a grassy margin. Studies comparing multiple flowering margins without a control that did not include spontaneous vegetation, grassy margins, or a crop were considered to lack a low-level plant diversity control and

were discarded. However, if a low diversity control was present, we included articles with multiple different high diversity treatments (which were grouped or separated depending on the analysis, see Section 2.2). (3) The article deals with arthropods (insects, arachnids, etc.). (4) The article or data available in Supporting Information presents data that can be extracted from text, tables, or figures. Reviews and meta-analyses were not considered. Articles only presenting results of inferential statistics, such as p -values, associated with tests between different diversification levels were not considered. (5) The article presents data from samplings conducted in the crop. Studies reporting, for example, only the abundance of arthropods in different SNH were not considered. (6) The article presents data from “agriculturally realistic” experiments. Data were not considered if the experiment was done on potted plants (except if this was a standard agricultural practice, for example, in horticulture), or if the experiment was done in controlled conditions, e.g., a laboratory or experimental glasshouses.

A total of 427 articles met these criteria and were analyzed carefully to extract data (see Fig. S1 for details of the reason for excluding articles). This dataset was supplemented with 22 articles obtained from the reference lists of other published meta-analyses conducted on agricultural diversification (Letourneau et al. 2011; Lichtenberg et al. 2017; Albrecht et al. 2020; Wan et al. 2020; Zamorano et al. 2020). Each article was then inspected and data extracted from text, figures, tables, and Supporting Information. Criteria were then applied to remove redundant values belonging to the same article (see Supporting Information). For each observation, the diversification practice used as well as information to characterize how the practice was implemented (e.g., management factors) were also recorded (see Table S1).

2.2 Statistical analysis

The data extracted from the articles comprised observations of the arthropod response (abundance of arthropods or provision of services/disservices) to different treatment types (non-diverse control and different diversification practices), alongside information about the response type (what kind of arthropods or services; Fig. 2). Our analysis aimed to determine how the response differed depending on both the treatment type and the response type; in other words, testing how different types of arthropods and services responded to different diversification practices. We took a hierarchical approach, partly inspired by Wan et al. (2020), with initial models providing a general overview of positive and negative effects of diversification, and subsequent models investigating associations between specific categories of diversification practice, arthropods, and services. First, we evaluated whether diversification in general

(all four practices grouped together) had an overall effect on two response groups: beneficial arthropods and ecosystem services (including predators, parasitoids, pollinators, predation, parasitism, and pollination) and antagonistic arthropods and ecosystem disservices (including herbivores and crop damage) (Model 1). Second, we tested whether the four diversification practices differed in their effects on beneficial arthropods and ecosystem services and on antagonists and disservices (Model 2). The third step was to test whether the diversification practices had different effects on different groups of arthropods and services/disservices (Model 3). Models 1–3 ran on our full dataset of 19,421 observations from 449 papers. Subsequent models tested the effect of different methods of implementing each diversification practice (Table S1) on the abundance of different arthropod groups and the provision of services/disservices (Models 4–7). For example, Model 4 compares control treatments without intercropping to intercrop treatments with different numbers of species, which were sown at different times, or in different layouts (Fig. 2, Table S1). Given that the implementation variables are nested within the relevant diversification practice, each of these models was run on the subset of data pertaining to each diversification practice.

For each model, we used the arm-based network meta-analysis method presented by Piepho et al. (2012, 2024). Arm-based meta-analysis involves extracting response data from each study for all treatments tested within that article (including the control), and then using a model to estimate the mean response for each treatment across all articles. Post hoc comparisons were then used to assess overall differences between each treatment and the control, and differences within response categories (e.g., in Model 1, comparisons were made between “non-diverse” and “diverse” within “beneficials and services” and within “antagonists and disservices”). This method differs from the traditional contrast-based meta-analysis method, where the differences between the treatments and the control are first calculated within each article (e.g., the log response ratio) and a model is used to estimate the mean difference across articles. The arm-based method is preferable for meta-analyses containing studies where the effects of multiple treatments are compared to a common control, because the method provides a better adjustment for within-study correlations between treatments, making it easier to split direct from indirect evidence (Piepho et al. 2024).

The response variable used in all our models was the log of the response recorded for each observation in each article. Taking the log allowed us to firstly correct for an increasing variability in the response as the mean response increased, and secondly to group together articles where the response was measured in different units and on different scales. The difference between two log-transformed values is the same as the log of their ratio, meaning that differences between

treatment means calculated on the log scale could be back-transformed to ratios, removing the units.

We implemented arm-based meta-analysis in R version 4.4.2 (R Core Team 2024) using the packages *glmmTMB* (Brooks et al. 2017) and *emmeans* (Lenth and Lenth 2018). The meta-analysis models were mixed effects linear models (which assumed a normal distribution and homoscedasticity of the residuals, after log transformation of the response). The treatment type and response type factors (Fig. 2), their interaction term, and the article identity were included as fixed effects. Interaction terms between each factor and article identity were included as random effects. The models were constructed using the *glmmTMB()* function, which enabled us to appropriately partition within vs between study variance by setting the “dispformula” argument to “~0 + obs,” where “obs” was a unique (categorical) identifier for every individual observation. The “start” and “map” arguments were used to set the starting value for each observation to the log of the inverse of the number of replicates that observation was based on. In meta-analysis, it is traditional to use the variance calculated by squaring the standard error (SE) for the observation extracted from the article. However, SEs can be calculated in a variety of ways across different studies, depending on the statistical techniques used. Furthermore, they are not always reported, and missing SEs have to be estimated. These differences in SE calculation methods can introduce bias into meta-analysis models, resulting in inaccurate mean estimates (regardless of whether they are arm-based or contrast-based). We found this to be a problem with our dataset and thus replaced the variance with the inverse of the number of replicates, which reduces the weighting given to observations derived from a small number of replicates. Where an article did not report the number of replicates, we conservatively assumed that the observation was derived from only one replicate.

Differences between treatments were assessed using the “~ctrl.vs.trt” method within the *contrasts()* function of *emmeans* to provide estimated treatment ratios (obtained by back-transforming differences between the log treatment means). The degrees of freedom were not correctly calculated for this type of model by *emmeans* (at the time of writing, it appeared the Kenward-Roger or Satterthwaite methods are not available for *glmmTMB* models in *emmeans*), so we manually adjusted these to the number of observations minus the number of fixed-effects parameters. Confidence intervals (95%) for the treatment ratios were also calculated by *emmeans*, which we used to determine whether the estimated ratio was significantly different from 1 (a ratio of 1 = no difference).

To assess potential publication bias in the dataset, we generated funnel plots by plotting precision ($1/\sqrt{\text{number of replicates}}$) against the effect size. Since the arm-based meta-analysis does not directly rely on effect size calculations, we

computed an effect size (log response ratio) for this purpose. For each study, this effect size was obtained by averaging the results of non-diversified and diversified treatments, considering the type of diversification practice, the classification of arthropods (beneficial vs antagonistic), their associated service or disservice, and the number of replicates. Separate funnel plots were then generated for each diversification practice and for each arthropod category (beneficial/service or antagonistic/disservice), and these plots were visually inspected to identify potential asymmetries indicative of publication bias.

3 Results and discussion

3.1 Studies published represent a good geographical coverage but are unequally distributed

The meta-analysis includes 19,421 observations (12,035 increased plant diversity treatments and 7386 non-diverse controls) from 449 studies. Intercropping comprises the majority of the diversified treatments (9234 observations, 289 studies), followed by flower resource addition (1821 observations, 106 studies), semi-natural habitat (SNH) (511 observations, 36 studies), and agroforestry (469 observations and 28 studies). In terms of response variables, most observations were on herbivore abundance (9721 observations from 302 studies), followed by predator abundance (4839 observations from 213 studies), parasitism rate (1523 observations from 88 studies), crop damage by herbivores (1408 observations from 88 studies), parasitoid abundance (1109 observations from 71 studies), pollinator abundance (347 observations from 44 studies), predation rate (421 observations from 37 studies), and finally pollination (53 observations from 14 studies).

Studies used for the analysis were published between January 1980 and June 2024, peaking in 2023 with 38 publications (Fig. 3B). For most response categories, studies started to be published on a frequent basis around the 1990s and increased over time, except for pollinators and pollination, where studies started to be published frequently only after the 2010s (Fig. 3C).

The included studies encompass all continents with cropped land (Fig. 3A): Europe (3235 observations from 129 studies), North America (4538 observations from 110 studies), Asia (5445 observations from 94 studies), Africa (3619 observations from 66 studies), South America (1683 observations from 33 studies), and Oceania (904 observations from 20). Intercropping is well represented in all continents, with a minimum of 12 studies found in Oceania and a maximum of 80 in Asia. However, the others are mainly found in Europe or North America, and in particular, SNH

is not represented in Africa or Oceania. This unbalanced geographic distribution of studies reflects past and current discrepancies in research conducted in different countries.

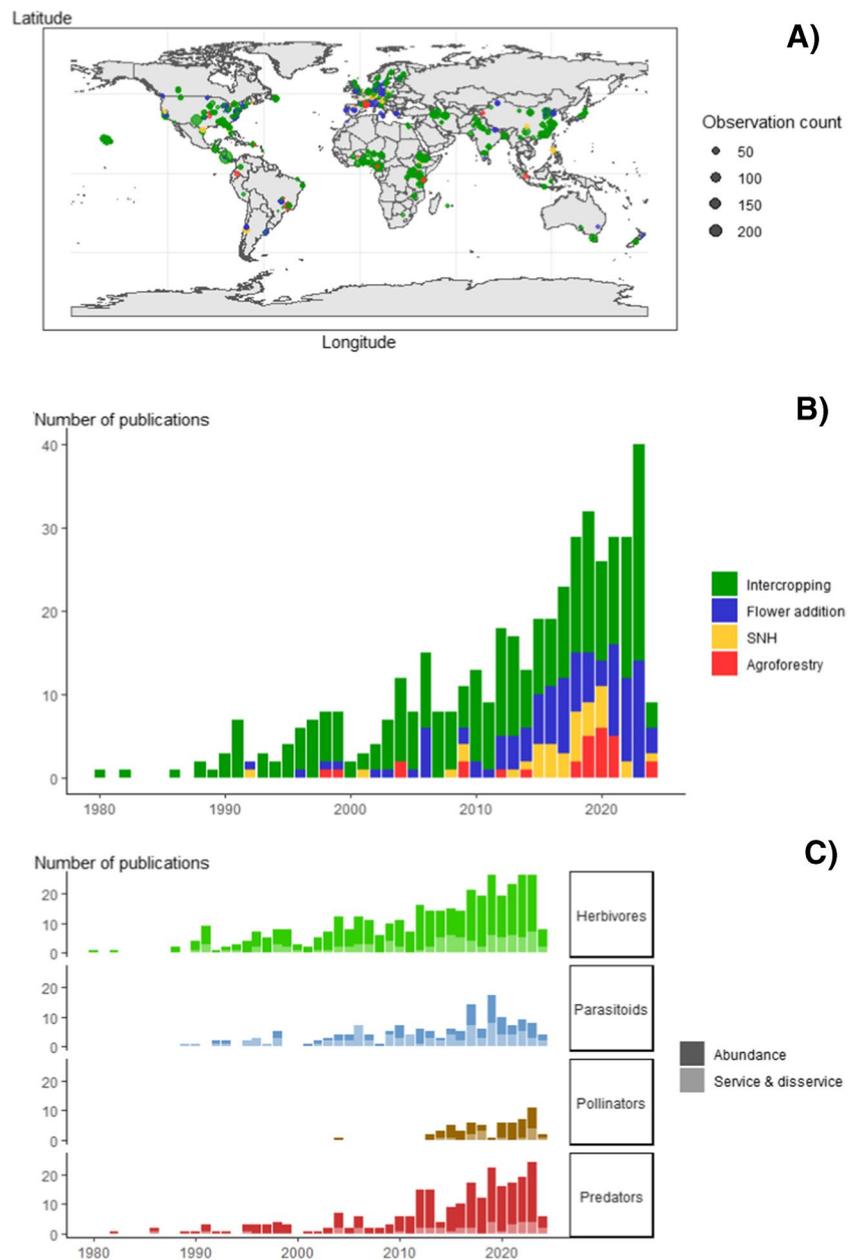
Funnel plots created to check for publication bias do not show major signs of publication bias for the different diversification practices and the classification of arthropods (beneficial vs antagonistic) and their associated service or disservice (Fig. S4).

3.2 Plant diversification has positive effects on ecosystem services and pest management

When comparing all diversification practices to non-diverse control practices, we observed an overall positive effect of diversification on beneficial arthropods (parasitoids, predators, and pollinators) and the ecosystem services they provide, leading to an average 41% (CI 95% 27–56%) increase, while reducing herbivore abundance and damage by an average of –33% (CI 95% –39 to –26%) (Fig. 4, Table S2). These results align with previous meta-analyses highlighting the positive effect of plant diversification practices, including agroforestry, addition of flower resources, intercropping, and maintenance of SNH (Lichtenberg et al. 2017; Wan et al. 2020, 2022; Tamburini et al. 2020; Beillouin et al. 2021). The observed benefit can be attributed to a combination of bottom-up and top-down mechanisms. Addition of flower resources, maintenance of SNH, and agroforestry are often implemented to increase the population of natural enemies in the crop, leading to higher ecosystem services that consequently negatively affect arthropod pest abundance and their damage to crops (Ratnadass et al. 2012). Intercropping can also lead to increased natural enemy populations by providing them with resources including nectar, pollen, and secondary prey. Intercropping is commonly implemented with mixtures of cereals and legumes (Kirsch et al. 2023) where legume flowers or extrafloral nectar can be used by some natural enemies for food. These species mixtures often also host populations of aphids used by predators as an alternative prey or a source of honeydew (Luquet et al. 2021). A direct effect of plant diversity on herbivores can also occur in intercropping, where practices such as trap-cropping and repellent-cropping disrupt herbivore colonization of crop plants (Shelton and Badenes-Perez 2006).

The effect of plant diversification on arthropods and their related ecosystem services is highly variable between studies, so although the overall trend is positive, a lot of data are needed to reliably detect a significant positive mean effect. This variability between studies can be observed in our results: the means for treatment and/or response categories with fewer observations from fewer studies have wider confidence intervals. This variability justifies the need to explore why diversification is more effective in some cases and less in others. In our subsequent analyses, we explore whether

Fig. 3 Map and temporal trend in the publication of the data used for the meta-analysis. **A** Map of the location of the different observations collected in the meta-analysis on the impact of plant diversification practices on arthropod abundance and ecosystem services. Duplicated locations were removed, and some observations could not be located because of a lack of information in the articles. **B** Number of publications used in the meta-analysis published per year between January 1980 and June 2024. Dots and bar colors represent different diversification practices: green, intercropping; blue, flower resource addition; yellow, semi-natural habitat (SNH); and red, agroforestry. **C** Number of publications used in the meta-analysis published per year, split between arthropod functional groups (herbivores, parasitoids, pollinators, and predators), with dark shade representing the abundance of arthropods and light shade the ecosystem services or disservices they cause.



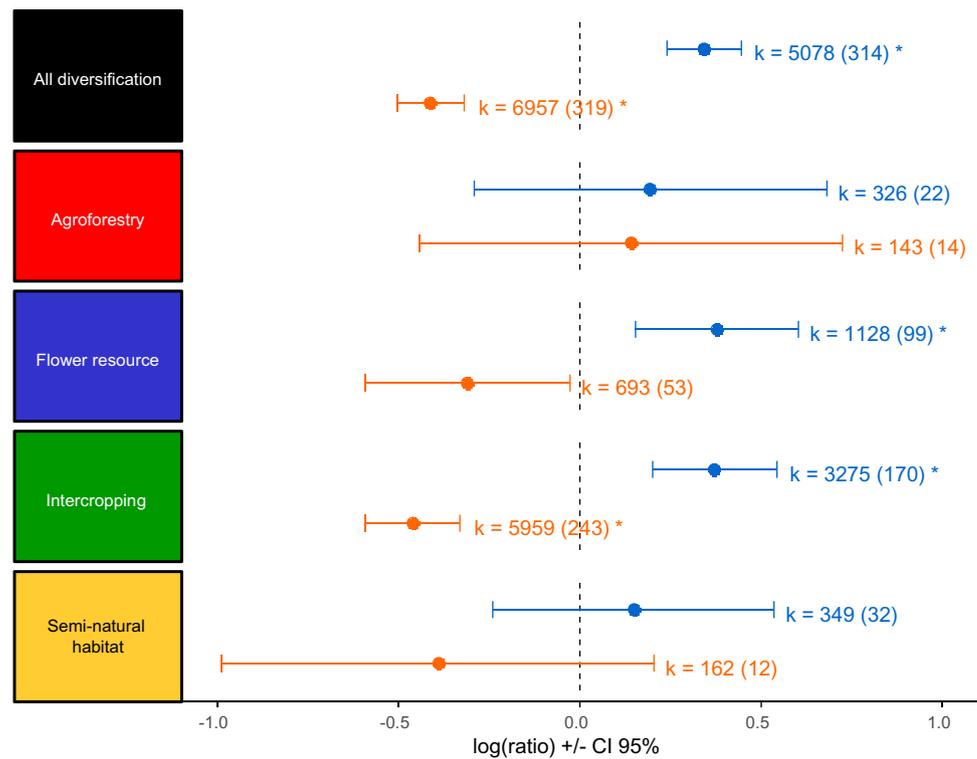
different arthropod groups and services respond differently to each diversification practice and whether the implementation and management of each diversification practice matter.

3.3 Diversification practices have different effects on arthropods and their services

Few meta-analyses have specifically compared how different diversification practices affect arthropod populations and the ecosystem services they provide to crops. Letourneau et al. (2011) distinguished between intercropping and flower addition, finding that both practices enhanced services to crops while reducing herbivore

pressure. Similarly, Albrecht et al. (2020) compared the effects of SNH and flower addition, showing a significant increase in pest control (predation and parasitism) with the addition of flowers alone. Our results concur with these findings: significant differences between monocrops and diversified plots are observed only for the addition of flower resources and the use of intercropping (Fig. 4, Table S2). These two practices lead to increases in the abundance of beneficial arthropod populations and their associated ecosystem services, and in the case of intercropping, to a reduction in herbivore abundance and crop damage. Intercropping and the addition of flower resources are the two practices for which we have the most

Fig. 4 Log of ratio (\pm CI 95%) of the effect of plant diversification practices on beneficial arthropods and ecosystem services (blue) and antagonists and disservices (orange) for all diversification practices and individual practices. k represents the number of observations in diversified treatments, and the numeral in brackets represents the number of publications per practice and response categories. Asterisk (*) highlights significant differences between control and diversified treatments.



data (Fig. S2), which might explain why the effects we observed are clearer than for other practices.

Beillouin et al. (2021) further divided diversification practices, including agroforestry, intercropping, cultivar mixtures, cover crops, and rotations, and demonstrated that both intercropping and agroforestry enhance pest and disease control. We did not find a positive result for agroforestry, but we have data from relatively few agroforestry studies (28) compared to intercropping (289) and flower resources (106), reducing the potential to detect a significant effect in the context of high between-study variability. Semi-natural habitats (with 35 studies) also do not yield statistically significant differences compared to monocrops, but follow a similar trend to intercropping and flower resources in a reduction of antagonists and an increase in beneficials (Fig. 4).

When examining the response of specific types of arthropods and specific services/disservices, significant effects were observed only when intercropping was implemented (Fig. 5, Table S2). Intercropping leads to a reduction of herbivore abundance by 39% (CI 95% 47–29%) compared to monocropping, a reduction of plant damage (–30% on average (CI 95% –46 to –8%)), and an increase in the abundance of predators and parasitoids on average of 48% (CI 95% 21–82%), and 56% (CI 95% 9–122%), respectively (Fig. 5). No significant effects of plant

diversification were detected for predation, parasitism, pollinator abundance, or pollination services. However, despite the lack of statistical significance, general trends for most of the practices indicate an increase in beneficial arthropod populations and ecosystem services, coupled with a reduction in herbivore populations and plant damage. Given the limited data availability for these categories, we are hesitant to conclude that the lack of significance indicates no effect across all categories; rather, it seems more plausible that there are small positive effects that are obscured by the high variability in outcomes between studies.

Other syntheses have also tested some of the categories shown in Fig. 5, finding similar results to ours. Specifically, Letourneau et al. (2011) also observed a reduction in herbivore abundance in intercropping systems, but not when flower margins are used. Albrecht et al. (2020) also failed to observe an effect of SNH and flower resource addition on pollination, but did observe a positive effect of flower addition on natural enemy abundance and parasitism rate. The latter is also hinted at in our data; the difference may relate either to the type of analyses used (Albrecht et al. 2020 were able to work with the raw data and thus account for more variability) or to the number of studies used (14 out of the 27 studies used in Albrecht et al. 2020 are common with the 106 studies included for SNH and flower addition in our analysis).

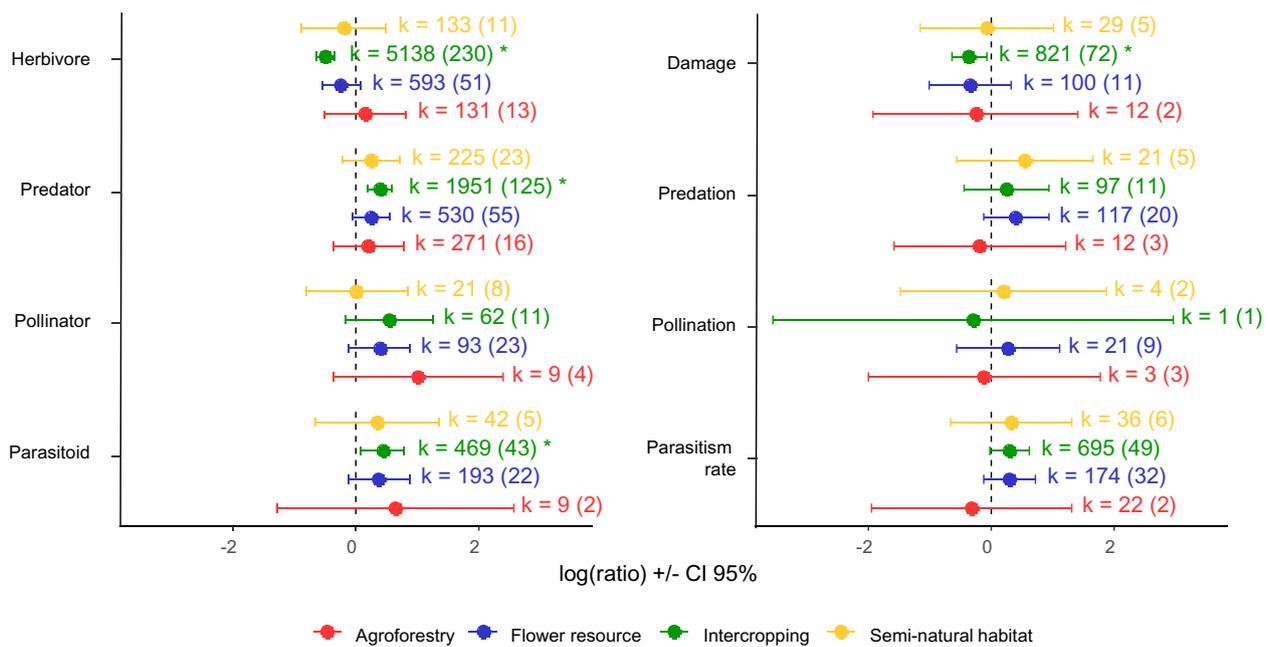


Fig. 5 Log of ratio (\pm CI 95%) of the effect for different response categories and different cropping practices. Green, intercropping; blue, flower resource addition; yellow, SNH; and red, agroforestry. Asterisk (*) highlights significant differences between control and diversified

treatments. k represents the number of observations in diversified treatments and in brackets the number of publications per practice and response categories.

3.4 The effect of management of diversification practices varies among different arthropods

In our investigation of the effects of management factors related to each diversification practice, we found significant but contrasting effects on herbivore abundance compared to parasitoid abundance and parasitism rate (Fig. 6, Table S2). Intercropping consistently suppressed herbivore abundance, while neither addition of flower resources nor SNH affected herbivores, regardless of how these diversification practices were managed. In contrast, parasitoids and parasitism were affected by management, with specific intercropping, flower resource, and SNH strategies leading to increased parasitism, and certain intercropping strategies leading to increased parasitoid abundances.

The link between the abundance and herbivore damage or parasitism was not clear. The general direction of effects between arthropods and services was consistent: intercropping reduced both herbivore abundance and damage, while usually increasing both parasitoids and parasitism. However, a significant effect on arthropod abundances was not usually associated with a significant effect on the associated service, and vice versa. This lack of relationship can be explained by the fact that other aspects of arthropod communities can impact the provision of services. Trait composition of communities can mediate this relationship, with examples of predator communities with smaller community weighted

mean body length being more effective at providing predation on aphids than larger ones (Rusch et al. 2015). Moreover, the total abundance of ecosystem service providers can hide interactions within this community, with intra-guild predation or competition, typically between predators and parasitoids, having potential negative effects on the delivery of the service (Frago 2016). Although almost all intercropping strategies significantly reduced herbivore abundance, crop damage was reduced under only two intercropping strategies. Similarly, parasitism rates were enhanced in several management categories of intercropping, addition of flower resources, and SNH that were not also found to enhance parasitoid abundance. This suggests that different factors may be important in regulating arthropod populations and their activity, so that one cannot simply assume that more or less of a given arthropod type will equate to higher or lower provision of their associated (dis-)service.

3.4.1 Intercropping

In intercropping, previous studies have shown that the spatial arrangement of crops can affect arthropods (Rakotomalala et al. 2023). Closer spacing between plant species can enhance the proximity of natural enemies to herbivores while simultaneously complicating herbivore host-plant location. Border cropping, a common strategy for trap-crops, involves sowing species that are more attractive to

		Herbivore	Damage	Predator	Predation	Parasitoid	Parasitism	Pollinator	Pollination
Agroforestry	Alley cropping	∅	∅	∅	∅	↗	∅	∅	∅
	Managed forest	∅	∅	∅	∅	∅	∅	∅	∅
	Fruits and nuts	∅	∅	∅	∅	∅	∅	∅	∅
	Diverse tree functions	∅	∅	∅	∅	∅	∅	∅	∅
Intercropping	Cash crops	↘	↘	∅	∅	↗	∅	∅	∅
	Companion crops	↘	∅	∅	∅	∅	∅	∅	∅
	Strip cropping	↘	∅	∅	∅	∅	↗	∅	∅
	Border cropping	↘	∅	∅	∅	∅	∅	∅	∅
	Mixed cropping	↘	∅	∅	∅	↗	∅	∅	∅
	2 crops	↘	∅	∅	∅	↗	∅	∅	∅
	3 crops	↘	∅	∅	∅	↗	↗	∅	∅
	>3 crops	↘	∅	∅	∅	∅	∅	∅	∅
	Sowing before	↘	∅	∅	∅	∅	∅	∅	∅
	Sowing simultaneously	↘	∅	∅	∅	∅	∅	∅	∅
Sowing after	∅	↘	∅	∅	∅	∅	∅	∅	
Flower resource	Separated from flowers	∅	∅	∅	∅	∅	↗	∅	∅
	Mixed with flowers	∅	∅	∅	∅	∅	∅	∅	∅
	< 5 species	∅	∅	∅	∅	∅	↗	∅	∅
	> 5 species	∅	∅	∅	∅	∅	∅	∅	∅
	Surface < 100m ²	∅	∅	∅	∅	∅	∅	∅	∅
	Surface > 100m ²	∅	∅	∅	∅	∅	∅	∅	∅
	1 year old	∅	∅	↗	∅	∅	∅	∅	∅
	>1 year old	∅	∅	∅	∅	∅	↗	∅	∅
SNH	< 10m from SNH	↗	∅	∅	∅	∅	∅	∅	∅
	> 10m from SNH	∅	∅	∅	∅	∅	∅	∅	∅
	Woody habitat	∅	∅	∅	∅	∅	↗	∅	∅
	Herbaceous habitat	∅	∅	∅	∅	∅	∅	∅	∅

Fig. 6 Summary of the effect of different management variables in studies using different plant diversification practices compared to monocrops on different response categories. Red arrows indicate a decrease of the response for a level of the variable tested, a green arrow indicates an increase, and a barred zero indicates that not

enough points were available to test this effect. Colored cells (red or green) show significant results. Explanations of each category are provided in Table S1, while details of the number of observations and studies for each level of the variables are presented in Table S3.

herbivores than the crop along crop edges to divert pests from the main crop (Shelton and Badenes-Perez 2006). This approach is expected to decrease herbivore abundance without significantly affecting natural enemies or pollinators. Our findings partially align with these predictions: increased parasitoid populations and parasitism rates were observed in configurations where crops were closely interspersed (e.g., strip and mixed cropping) but not in border cropping (Fig. 6, Fig. S5). Strong evidence supports a reduction in herbivore populations across all tested intercropping configurations, highlighting the effectiveness of this practice in decreasing herbivore pressure (Fig. 6, Fig. S5).

While a positive relationship between plant species diversity and ecosystem services is well-documented in

natural systems (Wan et al. 2022), this trend was not consistently observed in the intercropping systems analyzed. An increase in parasitoid abundance and parasitism rates was found when two or three crops were grown simultaneously, whereas reductions in herbivore populations occurred under all intercropping conditions (Fig. 6, Fig. S5). This may be because, in agricultural settings, plant species are often selected for specific functions (e.g., improving soil fertility, providing ground cover, or repelling pests). As a result, a well-designed system with fewer, strategically chosen species may equal or outperform a more diverse plant community. Push-pull systems illustrate this concept: they effectively manage pests and enhance parasitism services using just three species: a main crop, a pest-repellent crop,

and a pest-attractive crop planted on the field's perimeter (Cook et al. 2007).

Differences in sowing time between plant species in a field can lead to differences in plant biomass (Cotes et al. 2018). Plants with high biomass can more easily hide another, smaller crop plant from herbivores, and thus reduce abundance and damage. Our results partially support this premise, with reductions in herbivore abundance on the crop observed when the secondary crop is sown either before or at the same time as the main crop, but not after. However, contrary to expectations, a reduction in damage was only found when the secondary crop was sown after the main crop. We did not investigate whether it is more common to sow different types of intercrop at different times, so this could be an effect of specific crop combinations rather than the timing of sowing.

The use of companion plants (plants sown but not harvested) can be designed to provide services such as ground cover to reduce weed pressure but also to reduce herbivore pest abundance and damage (Lorin et al. 2015; Verret et al. 2017). However, the data available support the fact that herbivores and their damage are significantly reduced in systems with cash crops only, but not when companion crops are used. A significant increase in parasitoid abundance is also observed when only cash crops are used. A reason for this might be that companion plants are usually selected not to compete with the cash crop, and so potentially have less capacity to hide the cash crop from herbivores. Well-known examples of protective effects include ground-covering legumes as companion crops to oilseed rape, which are efficient in reducing herbivore pressure early in the season when plants are small, but their effect later in the season (when the oilseed rape is taller) is less clear (Seimandi-Corda et al. 2024; Magnin et al. 2025). In contrast, intercropped cash crops are more often of a similar stature, e.g., wheat and faba bean, barley and pea, and may therefore provide a stronger barrier effect.

3.4.2 Flower resources

Plant species diversity of flower margins, the size of the flower patches, the time since establishment of the patches, and the distance between the sampling location and the flowers can have an impact on arthropods. More diverse flower mixtures increase the complementarity of flower resource provision in space and time to better support beneficial arthropods (Scheper et al. 2013; Sutter et al. 2017). Plurianual flower margins are less disturbed and provide better shelter for overwintering beneficials than annual margins. Larger flower patches provide more resources which support arthropod populations, and the populations of beneficial arthropods are expected to be higher in flower margins than in the crop. Consequently, it is expected to have more

abundant beneficial populations close to flower patches than within the crop (Krimmer et al. 2019). Some of these effects are supported by Albrecht et al. (2020) for pollination services, but not for pest control. The present results show a more variable perspective. We found that parasitism rates increased significantly when flower resources were older than 1 year, but predator abundance increased significantly only when flower resources were younger than 1 year. Potentially, the soil disturbance involved in establishing flower resources in the first year, combined with the succession of the flower resource community over time, could favor different arthropod guilds at different times. Parasitism rates were also observed to be higher alongside less diverse flower resources (< 5 species) than in more diverse patches, and when samples were collected at least 1 m away from the flower resource into the crop. It is not clear what mechanisms could underlie these trends, so these results should be treated with caution until further research can shed more light on them. In particular, it is surprising that flower patch size had no effect on arthropods (Fig. 6, Fig. S5).

3.4.3 Semi-natural habitat (SNH)

It has been previously established that ecosystem service providers or pests can spill over from SNH to affect the crop (Rand et al. 2006; Boetzel et al. 2024). Our analysis indicates that such spillover effects may not always be positive, with an increase in herbivore abundance observed within 10 m of SNH. Some agricultural pests complete part of their development cycle in SNH, potentially explaining increased herbivore abundances near SNH (Juhel et al. 2017; Pigot et al. 2024). We also observed an increase in parasitism when woody SNH (hedgerows or woodland) was present, which may help to mitigate the effect of increased herbivore abundance (Fig. 6, Fig. S3). However, the data currently available on SNH remain limited, and additional studies are needed to confirm their effects. Landscape-level studies suggest that a higher proportion of grassland is associated with increased pollinator abundance, greater predation, or reduced pest pressure, while the proportions of forests and hedgerows appear to have weaker effects (Bartual et al. 2019; Perrot et al. 2023). To date, however, no meta-analysis has investigated how these different habitat types individually influence arthropods at the landscape scale.

3.4.4 Agroforestry

Few studies have tested the effect of agroforestry on arthropods, and thus, we have insufficient data to draw on to test the effects on predation, pollinators, and pollination. The only significant effect observed was the positive effect of alley cropping agroforestry on parasitoid abundance, which might reflect the fact that trees can provide refugia

to beneficial arthropods and thus benefit their populations. However, not enough studies are currently available to test whether the benefit of agroforestry on ecosystem service providers is better supported by different configurations of tree plantations. In managed forests where coffee and cacao are grown, a positive relationship between the tree species diversity and the abundance of parasitoids and pollinators, and reduced pest infestation has been observed (Sperber et al. 2004; Nesper et al. 2017; Geeraert et al. 2019); but it is not sure if this relationship can be applied to alley cropping system.

3.5 Limitations of the dataset and knowledge gaps

A key feature in our results is the high degree of between-study variability, leading to lower confidence in effects where less data is available. In some cases, it is therefore difficult to distinguish whether some practices or management factors truly have no effect or whether we do not have enough data to detect an effect. However, this variability itself is also informative, indicating that the effects are inconsistent. In that regard, our results support previous studies showing mixed effects of plant and habitat diversity on arthropods and their services (e.g., Karp et al. 2018). In separating the four diversification practices into different management strategies, we had hoped to explain some of this variation and to identify management strategies with more consistent effects than others. Although our results shed light on the effects of management practice (i.e., showing that all intercropping strategies reduce herbivores, while only some intercropping strategies increase parasitoids' abundance), it does not seem that management is an overarching driver of the effect of plant diversification practices on arthropods. We initially sought to account for the context dependency of these effects by incorporating additional variables such as abiotic factors (e.g., climate) and more detailed information on plant and arthropod traits or phylogenetic groups. However, insufficient data prevented us from reliably testing these factors. Future studies that directly compare multiple management practices will be valuable for clarifying the efficacy of diversification strategies, but careful consideration of context will remain essential for interpreting their outcomes.

The high degree of between-study variability in our data meant that we tend to see more significant effects in categories for which we have more data. Notably, less data is available for agroforestry and maintenance of SNH than for intercropping and flower addition (Fig. S3). These smaller datasets likely contribute fewer significant results regarding these practices. Agroforestry and SNH are often considered practices that are challenging to implement, requiring a long establishment time and complex management, which may partially explain why they are less studied. They have also

only recently gained substantial research attention (Fig. 3), while intercropping has been extensively studied, particularly for its agronomic benefits, for a much longer period. Flower resources have been recognized as a way to support biodiversity in agricultural landscapes (Jönsson et al. 2015), which may also explain why more studies are available here. However, another reason that we could not include many studies involving agroforestry and SNH is that they often compare plots along gradients of tree diversity or landscape coverage of SNH, which do not match our criteria for inclusion of the studies.

A lack of data is also observed for some types of responses, particularly for pollinator abundance and pollination service (Fig. 3). It highlights that even if crop diversification is a topic with an abundant literature (about 450 studies used in the present meta-analysis, and more studies that did not fit our criteria for inclusion), there is still a need for more research to be conducted. Interest in pollinators and their services is also relatively recent and is increasing with concerns about the negative impact of farming on populations of pollinators (Gemmill-Herren et al. 2021). It should also be noted that some service-providing arthropod groups, such as detritivores, are entirely missing from our study due to a lack of data. These organisms are important in nutrient cycles and for soil fertility, but are clearly understudied.

4 Conclusions

Our study presents one of the largest and most up-to-date meta-analyses of the effect of plant diversification on arthropods and their services in agriculture. It shows that plant diversification has the potential to enhance beneficial arthropod populations and the ecosystem services they provide, while reducing herbivorous pest populations and their associated crop damage. By comparing diversification practices, we showed for the first time that the strongest and most consistent effects were observed for intercropping, leading to significant increases in predator and parasitoid populations, and reductions in herbivore abundance and damage. However, only limited effects were observed for the addition of flower resources, agroforestry, and SNH, as well as for certain key ecosystem services like pollination. These gaps highlight the need for further research.

Implementing plant diversification practices can require a substantial redesign of farming systems, or farmer cooperation and local governance in the case of SNH, that are not always owned by neighboring farmers. These changes must be supported by institutions, farmer networks, and evidence-based knowledge. Differences in the way diversification practices are implemented can lead to complex effects on arthropods, which are rarely taken into consideration in meta-analyses. An originality of the present study

is to consider these management factors, showing that the effect of diversification practices is variable depending on how they are implemented and highly context-dependent. Consequently, while plant diversification shows clear potential to reduce pesticide reliance and support biodiversity, its practical implementation will require tailored strategies that account for local conditions. To promote the adoption of these practices, especially intercropping, policies that provide incentives and support for farmers are crucial. In the UK Sustainable Farming Incentive, intercropping is an option (<https://www.gov.uk>), but more advice as to effective combinations is necessary. While more research is needed to understand underexplored practices, there is sufficient evidence to justify action in favor of plant diversification, as overall these practices have the potential to pave the way to a more sustainable future food production.

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1007/s13593-025-01082-7>.

Authors' contributions Conceptualization, G.S.C., S.C., A.M.; methodology, C.M. and J.F.; formal analysis, G.S.C., C.M., J.H.; resources, G.S.C., K.T., A.D.; data curation, G.S.C.; writing—original draft, G.S.C., C.M., K.T.; writing—review and editing, A.M., S.C., J.F.; visualization, G.S.C., C.M., K.T.; supervision, A.M., S.C.; project administration, S.C.; funding acquisition, S.C.

Funding This work was funded by the European Union's Horizon 2020 Research and Innovation program as part of the project EcoStack (Grant Agreement no. 773554). Rothamsted Research receives strategic funding from the Biotechnology and Biological Sciences Research Council of the United Kingdom (BBSRC). SMC was part-funded by the Growing Health Institute Strategic Program (BB/X010953/1; BBS/E/RH/230003C). CM was partly supported by a Horizon Europe Marie Skłodowska-Curie Action Postdoctoral Fellowship (grant agreement 101063481, "EcoDiv").

Data availability Data used for the study can be found at <https://zenodo.org/records/15023242>.

Code availability R code used for the analysis is available in Supporting materials.

Declarations

Ethics approval Not applicable

Consent to participate Not applicable

Consent to publication Not applicable

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in

the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Aizen MA, Garibaldi LA, Cunningham SA, Klein AM (2009) How much does agriculture depend on pollinators? Lessons from long-term trends in crop production. *Ann Bot* 103:1579–1588
- Albrecht M, Kleijn D, Williams NM et al (2020) The effectiveness of flower strips and hedgerows on pest control, pollination services and crop yield: a quantitative synthesis. *Ecol Lett* 23:1488–1498. <https://doi.org/10.1111/ele.13576>
- Bartual AM, Sutter L, Bocci G et al (2019) The potential of different semi-natural habitats to sustain pollinators and natural enemies in European agricultural landscapes. *Agric Ecosyst Environ* 279:43–52
- Beillouin D, Ben-Ari T, Malézieux E et al (2021) Positive but variable effects of crop diversification on biodiversity and ecosystem services. *Glob Change Biol* 27:4697–4710. <https://doi.org/10.1111/gcb.15747>
- Blesh J, Mehrabi Z, Wittman H et al (2023) Against the odds: network and institutional pathways enabling agricultural diversification. *One Earth* 6:479–491. <https://doi.org/10.1016/j.oneear.2023.03.004>
- Boetzi FA, Sponsler D, Albrecht M et al (2024) Distance functions of carabids in crop fields depend on functional traits, crop type and adjacent habitat: a synthesis. *Proc R Soc Lond B Biol Sci* 291:20232383. <https://doi.org/10.1098/rspb.2023.2383>
- Bommarco R (2024) Ecological redesign of crop ecosystems for reliable crop protection. A review. *Agron Sustain Dev* 44:51. <https://doi.org/10.1007/s13593-024-00987-z>
- Brooker RW, Hawes C, Iannetta PPM et al (2023) Plant diversity and ecological intensification in crop production systems. *J Plant Ecol* 16:rtad015. <https://doi.org/10.1093/jpe/rtad015>
- Brooks ME, Kristensen K, Van Benthem KJ et al (2017) glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. *R J* 9:378–400
- Cook SM, Khan ZR, Pickett JA (2007) The use of push-pull strategies in integrated pest management. *Annu Rev Entomol* 52:375–400
- Cotes B, Rämert B, Nilsson U (2018) A first approach to pest management strategies using trap crops in organic carrot fields. *Crop Prot* 112:141–148. <https://doi.org/10.1016/j.cropro.2018.05.025>
- Crowther LI, Wilson K, Wilby A (2023) The impact of field margins on biological pest control: a meta-analysis. *Biocontrol* 68:387–396. <https://doi.org/10.1007/s10526-023-10205-6>
- Dainese M, Martin EA, Aizen MA et al (2019) A global synthesis reveals biodiversity-mediated benefits for crop production. *Sci Adv* 5:eaax0121. <https://doi.org/10.1126/sciadv.aax0121>
- Emmerson M, Morales MB, Oñate JJ, et al (2016) How agricultural intensification affects biodiversity and ecosystem services. In: *Advances in ecological research*. Elsevier, pp 43–97
- Frago E (2016) Interactions between parasitoids and higher order natural enemies: intraguild predation and hyperparasitoids. *Curr Opin Insect Sci* 14:81–86. <https://doi.org/10.1016/j.cois.2016.02.005>
- Geeraert L, Aerts R, Jordaens K et al (2019) Intensification of Ethiopian coffee agroforestry drives impoverishment of the Arabica coffee flower visiting bee and fly communities. *Agroforest Syst* 93:1729–1739. <https://doi.org/10.1007/s10457-018-0280-0>

- Gemmill-Herren B, Garibaldi LA, Kremen C, Ngo HT (2021) Building effective policies to conserve pollinators: translating knowledge into policy. *Curr Opin Insect Sci* 46:64–71
- Grassini P, Eskridge KM, Cassman KG (2013) Distinguishing between yield advances and yield plateaus in historical crop production trends. *Nat Commun* 4:2918. <https://doi.org/10.1038/ncomms3918>
- How to do the SFI actions for integrated pest management. In: GOV. UK. <https://www.gov.uk/guidance/how-to-do-the-sfi-actions-for-integrated-pest-management>. Accessed 4 Feb 2025
- Hufnagel J, Reckling M, Ewert F (2020) Diverse approaches to crop diversification in agricultural research. A review. *Agron Sustain Dev* 40:14. <https://doi.org/10.1007/s13593-020-00617-4>
- Isbell F, Adler PR, Eisenhauer N et al (2017) Benefits of increasing plant diversity in sustainable agroecosystems. *J Ecol* 105:871–879. <https://doi.org/10.1111/1365-2745.12789>
- Iverson AL, Marín LE, Ennis KK et al (2014) Review: do polycultures promote win-wins or trade-offs in agricultural ecosystem services? A meta-analysis. *J Appl Ecol* 51:1593–1602. <https://doi.org/10.1111/1365-2664.12334>
- Jachowicz N, Sigsgaard L (2025) Highly diverse flower strips promote natural enemies more in annual field crops: a review and meta-analysis. *Agric Ecosyst Environ* 381:109412. <https://doi.org/10.1016/j.agee.2024.109412>
- Jaworski CC, Thomine E, Rusch A et al (2023) Crop diversification to promote arthropod pest management: a review. *Agriculture Communications* 1(1):100004. <https://doi.org/10.1016/j.agr-com.2023.100004>
- Jönsson AM, Ekroos J, Dänhardt J et al (2015) Sown flower strips in southern Sweden increase abundances of wild bees and hoverflies in the wider landscape. *Biol Conserv* 184:51–58
- Juhel AS, Barbu CM, Franck P et al (2017) Characterization of the pollen beetle, *Brassicogethes aeneus*, dispersal from woodlands to winter oilseed rape fields. *PLoS ONE* 12:e0183878. <https://doi.org/10.1371/journal.pone.0183878>
- Karp DS, Chaplin-Kramer R, Meehan TD, et al (2018) Crop pests and predators exhibit inconsistent responses to surrounding landscape composition. *Proc Natl Acad Sci* 115:. <https://doi.org/10.1073/pnas.1800042115>
- Kass M (2020) Summary for policymakers of the global assessment report on biodiversity and ecosystem services. *Nat Resour Environ* 34(3):62–62
- Kirsch F, Hass AL, Link W, Westphal C (2023) Intercrops as foraging habitats for bees: bees do not prefer sole legume crops over legume-cereal mixtures. *Agric Ecosyst Environ* 343:108268. <https://doi.org/10.1016/j.agee.2022.108268>
- Kremen C, Iles A, Bacon C (2012) Diversified farming systems: an agroecological, systems-based alternative to modern industrial agriculture. *Ecol Soc* 17:art44. <https://doi.org/10.5751/ES-05103-170444>
- Krimmer E, Martin EA, Krauss J et al (2019) Size, age and surrounding semi-natural habitats modulate the effectiveness of flower-rich agri-environment schemes to promote pollinator visitation in crop fields. *Agric Ecosyst Environ* 284:106590. <https://doi.org/10.1016/j.agee.2019.106590>
- Lenth R, Lenth MR (2018) Package ‘lsmeans.’ *Am Stat* 34:216–221
- Letourneau DK, Armbrrecht I, Rivera BS et al (2011) Does plant diversity benefit agroecosystems? A synthetic review. *Ecol Appl* 21:9–21. <https://doi.org/10.1890/09-2026.1>
- Lichtenberg EM, Kennedy CM, Kremen C et al (2017) A global synthesis of the effects of diversified farming systems on arthropod diversity within fields and across agricultural landscapes. *Glob Change Biol* 23:4946–4957. <https://doi.org/10.1111/gcb.13714>
- Lorin M, Jeuffroy M-H, Butier A, Valantin-Morison M (2015) Undersowing winter oilseed rape with frost-sensitive legume living mulches to improve weed control. *Eur J Agron* 71:96–105. <https://doi.org/10.1016/j.eja.2015.09.001>
- Luquet M, Peñalver-Cruz A, Satour P et al (2021) Aphid honeydew may be the predominant sugar source for *Aphidius* parasitoids even in nectar-providing intercrops. *Biol Control* 158:104596. <https://doi.org/10.1016/j.biocontrol.2021.104596>
- Magnin L, Hiltbold I, Jullien A, Baux A (2025) Intercropping mitigates incidence of the oilseed rape insect pest complex. *Pest Manag Sci* ps.8835. <https://doi.org/10.1002/ps.8835>
- Nesper M, Kueffer C, Krishnan S et al (2017) Shade tree diversity enhances coffee production and quality in agroforestry systems in the Western Ghats. *Agric Ecosyst Environ* 247:172–181. <https://doi.org/10.1016/j.agee.2017.06.024>
- Nicholls CI, Altieri MA (2013) Plant biodiversity enhances bees and other insect pollinators in agroecosystems. A review. *Agron Sustain Dev* 33:257–274. <https://doi.org/10.1007/s13593-012-0092-y>
- Oerke E-C (2006) Crop losses to pests. *J Agric Sci* 144:31–43. <https://doi.org/10.1017/S0021859605005708>
- Perez-Alvarez R, Chara J, Snyder LD, et al (2023) Global meta-analysis reveals overall benefits of silvopastoral systems for biodiversity. *bioRxiv* 2023–07
- Perrot T, Rusch A, Gaba S, Bretagnolle V (2023) Both long-term grasslands and crop diversity are needed to limit pest and weed infestations in agricultural landscapes. *Proc Natl Acad Sci* 120:e2300861120. <https://doi.org/10.1073/pnas.2300861120>
- Piepho H-P, Forkman J, Malik WA (2024) A REML method for the evidence-splitting model in network meta-analysis. *Res Synth Methods* 15:198–212. <https://doi.org/10.1002/jrsm.1679>
- Piepho HP, Williams ER, Madden LV (2012) The use of two-way linear mixed models in multitreatment meta-analysis. *Biometrics* 68:1269–1277. <https://doi.org/10.1111/j.1541-0420.2012.01786.x>
- Pigot J, Gardarin A, Doré T et al (2024) Unlike woodland edges, flower strips do not act as a refuge for cabbage stem flea beetle aestivation. *Pest Manag Sci* 80:2325–2332. <https://doi.org/10.1002/ps.7558>
- Pumariño L, Sileshi GW, Gripenberg S et al (2015) Effects of agroforestry on pest, disease and weed control: a meta-analysis. *Basic Appl Ecol* 16:573–582
- R Core Team (2024) R: a language and environment for statistical computing
- Rakotomalala AANA, Ficiciyan AM, Tscharnke T (2023) Intercropping enhances beneficial arthropods and controls pests: a systematic review and meta-analysis. *Agric Ecosyst Environ* 356:108617. <https://doi.org/10.1016/j.agee.2023.108617>
- Rand TA, Tylianakis JM, Tscharnke T (2006) Spillover edge effects: the dispersal of agriculturally subsidized insect natural enemies into adjacent natural habitats. *Ecol Lett* 9:603–614. <https://doi.org/10.1111/j.1461-0248.2006.00911.x>
- Ratnadass A, Fernandes P, Avelino J, Habib R (2012) Plant species diversity for sustainable management of crop pests and diseases in agroecosystems: a review. *Agron Sustain Dev* 32:273–303. <https://doi.org/10.1007/s13593-011-0022-4>
- Raven PH, Wagner DL (2021) Agricultural intensification and climate change are rapidly decreasing insect biodiversity. *Proc Natl Acad Sci* 118:e2002548117. <https://doi.org/10.1073/pnas.2002548117>
- Rusch A, Birkhofer K, Bommarco R et al (2015) Predator body sizes and habitat preferences predict predation rates in an agroecosystem. *Basic Appl Ecol* 16:250–259. <https://doi.org/10.1016/j.baae.2015.02.003>
- Scheper J, Holzschuh A, Kuussaari M et al (2013) Environmental factors driving the effectiveness of European agri-environmental measures in mitigating pollinator loss – a meta-analysis. *Ecol Lett* 16:912–920. <https://doi.org/10.1111/ele.12128>
- Seimandi-Corda G, Winkler J, Jenkins T et al (2024) Companion plants and straw mulch reduce cabbage stem flea beetle (*Psylliodes chrysocephala*) damage on oilseed rape. *Pest Manag Sci* 80(5):2333–2341

- Shelton A, Badenes-Perez F (2006) Concepts and applications of trap cropping in pest management. *Annu Rev Entomol* 51:285–308
- Sperber CF, Nakayama K, Valverde MJ, Neves S (2004) Tree species richness and density affect parasitoid diversity in cacao agroforestry. *Basic Appl Ecol* 5:241–251. <https://doi.org/10.1016/j.baae.2004.04.001>
- Sutter L, Jeanneret P, Bartual AM et al (2017) Enhancing plant diversity in agricultural landscapes promotes both rare bees and dominant crop-pollinating bees through complementary increase in key floral resources. *J Appl Ecol* 54:1856–1864. <https://doi.org/10.1111/1365-2664.12907>
- Tamburini G, Bommarco R, Wanger TC et al (2020) Agricultural diversification promotes multiple ecosystem services without compromising yield. *Sci Adv* 6:eaba1715. <https://doi.org/10.1126/sciadv.aba1715>
- Verret V, Gardarin A, Makowski D et al (2017) Assessment of the benefits of frost-sensitive companion plants in winter rapeseed. *Eur J Agron* 91:93–103. <https://doi.org/10.1016/j.eja.2017.09.006>
- Wan N-F, Fu L, Dainese M et al (2022) Plant genetic diversity affects multiple trophic levels and trophic interactions. *Nat Commun* 13:7312. <https://doi.org/10.1038/s41467-022-35087-7>
- Wan N-F, Zheng X-R, Fu L-W et al (2020a) Global synthesis of effects of plant species diversity on trophic groups and interactions. *Nat Plants* 6:503–510. <https://doi.org/10.1038/s41477-020-0654-y>
- Zamorano J, Bartomeus I, Grez AA, Garibaldi LA (2020) Field margin floral enhancements increase pollinator diversity at the field edge but show no consistent spillover into the crop field: a meta-analysis. *Insect Conserv Divers* 13:519–531. <https://doi.org/10.1111/icad.12454>
- Alford L, Roudine S, Valsami D et al (2024) No evidence for competition over floral resources between winter-active parasitoids and pollinators in agroecosystems. *Sci Rep* 14:2239
- Alghali AM (1993) Intercropping as a component in insect pest management for grain cowpea, *Vigna unguiculata* Walp production in Nigeria. *Int J Trop Insect Sci* 14:49–54. <https://doi.org/10.1017/S1742758400013382>
- Amoabeng BW, Stevenson PC, Mochiah MB et al (2021) Economic analysis of habitat manipulation in *Brassica* pest management: wild plant species suppress cabbage webworm. *Crop Prot* 150:105788. <https://doi.org/10.1016/j.cropro.2021.105788>
- Ampong-Nyarko K, Reddy KVS, Nyang'or RA, Saxena KN (1994) Reduction of insect pest attack on sorghum and cowpea by intercropping. *Entomol Exp Appl* 70:179–184. <https://doi.org/10.1111/j.1570-7458.1994.tb00745.x>
- Armstrong G, McKinlay RG (1997) Vegetation management in organic cabbages and pitfall catches of carabid beetles. *Agric Ecosyst Environ* 64:267–276. [https://doi.org/10.1016/S0167-8809\(97\)00024-8](https://doi.org/10.1016/S0167-8809(97)00024-8)
- Arshad M, Ahmad S, Sufyan M, et al (2018) Population dynamics of aphids and their natural enemies associated with strip-intercropping in Wheat Crop. *Pak J Zool* 50. <https://doi.org/10.17582/journal.pjz/2018.50.4.1225.1230>
- Åsman K (2002) Trap cropping effect on oviposition behaviour of the leek moth *Acrolepiopsis assectella* and the diamondback moth *Plutella xylostella*. *Entomol Exp Appl* 105:153–164. <https://doi.org/10.1046/j.1570-7458.2002.01043.x>
- Asmare D, Muluken G, Seid H et al (2022) The effect of *Brachiaria* rows on stem borer damage on sorghum in eastern Amhara, Ethiopia. *Int J Trop Insect Sci* 42:2065–2071. <https://doi.org/10.1007/s42690-021-00637-x>
- Azandémè-Hounmalon GY, Logbo J, Gbèblonouo Dassou A et al (2023) Investigation of amaranth production constraints and pest infestation reduction by basil intercropping. *J Agric Food Res* 12:100627. <https://doi.org/10.1016/j.jafr.2023.100627>
- Azpiazu C, Medina P, Adán Á et al (2020) The role of annual flowering plant strips on a melon crop in central Spain. Influence on pollinators and crop. *Insects* 11:66. <https://doi.org/10.3390/insects11010066>
- Badenes-Pérez FR, Márquez BP, Petitpierre E (2017) Can flowering *Barbarea* spp. (Brassicaceae) be used simultaneously as a trap crop and in conservation biological control? *J Pest Sci* 90:623–633. <https://doi.org/10.1007/s10340-016-0815-y>
- Balmer O, Géneau CE, Belz E et al (2014) Wildflower companion plants increase pest parasitation and yield in cabbage fields: experimental demonstration and call for caution. *Biol Control* 76:19–27. <https://doi.org/10.1016/j.biocontrol.2014.04.008>
- Balmer O, Piffner L, Schied J et al (2013) Noncrop flowering plants restore top-down herbivore control in agricultural fields. *Ecol Evol* 3:2634–2646. <https://doi.org/10.1002/ece3.658>
- Balzan MV (2017) Flowering banker plants for the delivery of multiple agroecosystem services. *Arthropod-Plant Interact* 11:743–754. <https://doi.org/10.1007/s11829-017-9544-2>
- Balzan MV, Bocci G, Moonen A (2016) Utilisation of plant functional diversity in wildflower strips for the delivery of multiple agroecosystem services. *Entomol Exp Appl* 158:304–319. <https://doi.org/10.1111/eea.12403>
- Balzan MV, Moonen A (2014) Field margin vegetation enhances biological control and crop damage suppression from multiple pests in organic tomato fields. *Entomol Exp Appl* 150:45–65. <https://doi.org/10.1111/eea.12142>
- Barari H, Cook SM, Clark SJ, Williams IH (2005) Effect of a turnip rape (*Brassica rapa*) trap crop on stem-mining pests and their parasitoids in winter oilseed rape (*Brassica napus*). *Biocontrol* 50:69–86. <https://doi.org/10.1007/s10526-004-0895-0>

Reference of the meta-analysis

- Abad MKR, Fathi SAA, Nouri-Ganbalani G, Amiri-Besheli B (2020) Influence of tomato/clover intercropping on the control of *Helicoverpa armigera* (Hübner). *Int J Trop Insect Sci* 40:39–48. <https://doi.org/10.1007/s42690-019-00048-z>
- Adati T, Susila W, Sumiartha K et al (2011) Effects of mixed cropping on population densities and parasitism rates of the diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae). *Appl Entomol Zool* 46:247–253. <https://doi.org/10.1007/s13355-011-0036-z>
- Adeleye VO, Seal DR, Liburd OE et al (2022) Pepper weevil, *Anthonomus eugenii* (Coleoptera: Curculionidae) suppression on jalapeño pepper using non-host insect repellent plants. *Crop Prot* 154:105893. <https://doi.org/10.1016/j.cropro.2021.105893>
- Adhikari A, Reddy GVP (2017) Evaluation of trap crops for the management of wireworms in spring wheat in Montana. *Arthropod-Plant Interact* 11:755–766. <https://doi.org/10.1007/s11829-017-9533-5>
- Adler LS, Hazzard RV (2009) Comparison of perimeter trap crop varieties: effects on herbivory, pollination, and yield in butternut squash. *Environ Entomol* 38:207–215. <https://doi.org/10.1603/022.038.0126>
- Agboka K, Gounou S, Tamo M (2006) The role of maize-legumes-cassava intercropping in the management of maize ear borers with special reference to *Mussidia nigrivenella* Ragonot (Lepidoptera: Pyralidae). *Ann Soc Entomol Fr* 42:495–502. <https://doi.org/10.1080/00379271.2006.10697484>
- Ahmed S, Khan MA, Qasam M (2013) Effect of intercropping of maize in citrus orchards on citrus leaf miner infestation and population of its natural enemies. *Pak J Agri Sci* 50:91–93
- Alarcón-Segura V, Grass I, Breustedt G et al (2022) Strip intercropping of wheat and oilseed rape enhances biodiversity and biological pest control in a conventionally managed farm scenario. *J Appl Ecol* 59:1513–1523. <https://doi.org/10.1111/1365-2664.14161>

- Barbir J, Badenes-Pérez FR, Fernández-Quintanilla C, Dorado J (2015) Can floral field margins improve pollination and seed production in coriander *Coriandrum sativum* L. (Apiaceae)? can field margins improve pollination of coriander? *Agric for Entomol* 17:302–308. <https://doi.org/10.1111/afe.12108>
- Beaumelle L, Auriol A, Grasset M et al (2021) Benefits of increased cover crop diversity for predators and biological pest control depend on the landscape context. *Ecol Solutions Evidence* 2:e12086. <https://doi.org/10.1002/2688-8319.12086>
- Begum M, Gurr GM, Wratten SD et al (2006) Using selective food plants to maximize biological control of vineyard pests. *J Appl Ecol* 43:547–554. <https://doi.org/10.1111/j.1365-2664.2006.01168.x>
- Beizhou S, Jie Z, Jinghui H et al (2011) Temporal dynamics of the arthropod community in pear orchards intercropped with aromatic plants. *Pest Manage Sci* 67:1107–1114. <https://doi.org/10.1002/ps.2156>
- Beizhou S, Jie Z, Wiggins NL et al (2012) Intercropping with aromatic plants decreases herbivore abundance, species richness, and shifts arthropod community trophic structure. *Environ Entomol* 41:872–879. <https://doi.org/10.1603/EN12053>
- Belay D, Foster JE (2010) Efficacies of habitat management techniques in managing maize stem borers in Ethiopia. *Crop Prot* 29:422–428. <https://doi.org/10.1016/j.cropro.2009.09.006>
- Belay D, Schulthess F, Omwega C (2009) The profitability of maize–haricot bean intercropping techniques to control maize stem borers under low pest densities in Ethiopia. *Phytoparasitica* 37:43–50. <https://doi.org/10.1007/s12600-008-0002-7>
- Bell VA, Brightwell RJ, Lester PJ (2006) Increasing vineyard floral resources may not enhance localised biological control of the leafroller *Epiphyas postvittana* (Lepidoptera: Tortricidae) by *Dolichogenidea* spp. (Hymenoptera: Braconidae) parasitoids. *Biocontrol Sci Technol* 16:1031–1042. <https://doi.org/10.1080/09583150600828502>
- Bencharki Y, Christmann S, Lhomme P et al (2023) Farming with alternative pollinators' approach supports diverse and abundant pollinator community in melon fields in a semi-arid landscape. *Renew Agric Food Syst* 38:e6. <https://doi.org/10.1017/S1742170522000394>
- Bender DA, Morrison WP, Frisbie RE (1999) Intercropping cabbage and Indian mustard for potential control of lepidopterous and other insects. *HortSci* 34:275–279. <https://doi.org/10.21273/HORTSCI.34.2.275>
- Bensen TA, Temple SR (2008) Trap cropping, planting date, and cowpea variety as potential elements of an integrated pest management strategy for *Lygus hesperus* in blackeyed cowpea. *Crop Prot* 27:1343–1353. <https://doi.org/10.1016/j.cropro.2008.05.002>
- Berndt LA, Wratten SD, Hassan PG (2002) Effects of buckwheat flowers on leafroller (Lepidoptera: Tortricidae) parasitoids in a New Zealand vineyard. *Agric for Entomol* 4:39–45. <https://doi.org/10.1046/j.1461-9563.2002.00126.x>
- Berndt LA, Wratten SD, Scarratt SL (2006) The influence of floral resource subsidies on parasitism rates of leafrollers (Lepidoptera: Tortricidae) in New Zealand vineyards. *Biol Control* 37:50–55. <https://doi.org/10.1016/j.biocontrol.2005.12.005>
- Bhuiya MI, Rahman MA, Miah MMU et al (2020) Gliricidia tree leaf incorporation into soil and use of companion plants for safe tomato production. *J Fac Agric Kyushu Univ* 65:1–7. <https://doi.org/10.5109/2558881>
- Bickerton MW, Hamilton GC (2012) Effects of Intercropping With Flowering Plants on Predation of *Ostrinia nubilalis* (Lepidoptera: Crambidae) Eggs by Generalist Predators in Bell Peppers. *Environ Entomol* 41:612–620. <https://doi.org/10.1603/EN11249>
- Bigger DS, Chaney WE (1998) Effects of *Iberis umbellata* (Brassicaceae) on insect pests of cabbage and on potential biological control agents. *Environ Entomol* 27:161–167
- Bischoff A, Pollier A, Tricault Y et al (2022) A multi-site experiment to test biocontrol effects of wildflower strips in different French climate zones. *Basic Appl Ecol* 62:33–44. <https://doi.org/10.1016/j.baae.2022.04.003>
- Bishop GA, Fijen TPM, Desposato BN et al (2023) Hedgerows have contrasting effects on pollinators and natural enemies and limited spillover effects on apple production. *Agric Ecosyst Environ* 346:108364. <https://doi.org/10.1016/j.agee.2023.108364>
- Björkman M, Hambäck PA, Hopkins RJ, Rämert B (2010) Evaluating the enemies hypothesis in a clover-cabbage intercrop: effects of generalist and specialist natural enemies on the turnip root fly (*Delia floralis*). *Agric for Entomol* 12:123–132. <https://doi.org/10.1111/j.1461-9563.2009.00452.x>
- Björkman M, Hambäck PA, Rämert B (2007) Neighbouring monocultures enhance the effect of intercropping on the turnip root fly (*Delia floralis*). *Entomol Exp Appl* 124:319–326. <https://doi.org/10.1111/j.1570-7458.2007.00589.x>
- Blaauw BR, Isaacs R (2014) Flower plantings increase wild bee abundance and the pollination services provided to a pollination-dependent crop. *J Appl Ecol* 51:890–898. <https://doi.org/10.1111/1365-2664.12257>
- Blaauw BR, Isaacs R (2015) Wildflower plantings enhance the abundance of natural enemies and their services in adjacent blueberry fields. *Biol Control* 91:94–103. <https://doi.org/10.1016/j.biocontrol.2015.08.003>
- Blaauw BR, Morrison WR, Mathews C et al (2017) Measuring host plant selection and retention of *Halyomorpha halys* by a trap crop. *Entomol Exp Appl* 163:197–208. <https://doi.org/10.1111/eea.12571>
- Boetzel FA, Krimmer E, Krauss J, Steffan-Dewenter I (2019) Agri-environmental schemes promote ground-dwelling predators in adjacent oilseed rape fields: diversity, species traits and distance-decay functions. *J Appl Ecol* 56:10–20. <https://doi.org/10.1111/1365-2664.13162>
- Boinot S, Mézière D, Poulmarc'h J, et al (2020) Promoting generalist predators of crop pests in alley cropping agroforestry fields: Farming system matters. *Ecol Eng* 158:106041. <https://doi.org/10.1016/j.ecoleng.2020.106041>
- Booij C, Noorlander J, Theunissen J (1997) Intercropping cabbage with clover: effects on ground beetles. *Biol Agric Hortic* 15:261–268
- Bottenberg H, Tamò M, Singh BB (1998) Occurrence of phytophagous insects on wild *Vigna* sp. and cultivated cowpea: comparing the relative importance of host-plant resistance and millet intercropping. *Agric Ecosyst Environ* 70:217–229. [https://doi.org/10.1016/S0167-8809\(98\)00156-X](https://doi.org/10.1016/S0167-8809(98)00156-X)
- Boucher TJ, Ashley R, Durgy R et al (2003) Managing the pepper maggot (Diptera: Tephritidae) using perimeter trap cropping. *J Econ Entomol*. <https://doi.org/10.1093/jee/96.2.420>
- Brandmeier J, Reininghaus H, Pappagallo S et al (2021) Intercropping in high input agriculture supports arthropod diversity without risking significant yield losses. *Basic Appl Ecol* 53:26–38. <https://doi.org/10.1016/j.baae.2021.02.011>
- Brandmeier J, Reininghaus H, Scherber C (2022) Multi-species crop mixtures increase insect biodiversity in an intercropping experiment. *Authorea Preprints*
- Brandsæter LO, Netland J, Meadow R (1998) Yields, weeds, pests and soil nitrogen in a white cabbage-living mulch system. *Biol Agric Hortic* 16:291–309. <https://doi.org/10.1080/01448765.1998.10823201>
- Breitenmoser S, Steinger T, Baux A, Hiltbold I (2022) Intercropping winter oilseed rape (*Brassica napus* L.) has the potential to lessen the impact of the insect pest complex. *Agronomy* 12:723. <https://doi.org/10.3390/agronomy12030723>
- Broad ST, Schellhorn NA, Lissos SN et al (2008a) Host location and parasitism of *Brevicoryne brassicae* in diversified broccoli cropping systems. *Entomol Exp Appl* 129:166–171. <https://doi.org/10.1111/j.1570-7458.2008.00762.x>

- Broad ST, Schellhorn NA, Lisson SN, Mendham NJ (2008b) Host location and oviposition of lepidopteran herbivores in diversified broccoli cropping systems. *Agric for Entomol* 10:157–165. <https://doi.org/10.1111/j.1461-9563.2008.00374.x>
- Brown MW (2012) Role of biodiversity in integrated fruit production in eastern North American orchards. *Agric for Entomol* 14:89–99. <https://doi.org/10.1111/j.1461-9563.2011.00540.x>
- Brown MW, Mathews CR (2008) Conservation biological control of spirea aphid, *Aphis spiraeicola* (Hemiptera: Aphididae) on apple by providing natural alternative food resources. *Eur J Entomol* 105:537–540. <https://doi.org/10.14411/eje.2008.071>
- Brown MW, Mathews CR, Krawczyk G (2010) Extrafloral nectar in an Apple ecosystem to enhance biological control. *J Econ Entomol* 103:1657–1664. <https://doi.org/10.1603/EC10019>
- Bruce D, Silva EM, Dawson JC (2022) Suppression of weed and insect populations by living cover crop mulches in organic squash production. *Front Sustain Food Syst* 6:995224. <https://doi.org/10.3389/fsufs.2022.995224>
- Brust GE, Stinneri BR, McCartney DA (1986) Predation by soil inhabiting arthropods in intercropped and monoculture agroecosystems. *Agric Ecosyst Environ* 18:145–154. [https://doi.org/10.1016/0167-8809\(86\)90137-4](https://doi.org/10.1016/0167-8809(86)90137-4)
- Buchanan A, Grieshop M, Szendrei Z (2018) Assessing annual and perennial flowering plants for biological control in asparagus. *Biol Control* 127:1–8. <https://doi.org/10.1016/j.biocontrol.2018.08.013>
- Bugg RL, Wäckers FL, Brunson KE et al (1991) Cool-season cover crops relay intercropped with cantaloupe: influence on a generalist predator, *Geocoris punctipes* (Hemiptera: Lygaeidae). *J Econ Entomol* 84:408–416. <https://doi.org/10.1093/jee/84.2.408>
- Bui TNT, Mofikoya A, Blande JD et al (2024) Intercropping organic broccoli with *Rhododendron tomentosum* and *Fagopyrum esculentum*: a test of bottom-up and top-down strategies for reducing herbivory. *Arthropod-Plant Interact*. <https://doi.org/10.1007/s11829-023-10033-6>
- Bukovinszky T, Tréfas H, Van Lenteren JC et al (2004) Plant competition in pest-suppressive intercropping systems complicates evaluation of herbivore responses. *Agric Ecosyst Environ* 102:185–196. <https://doi.org/10.1016/j.agee.2003.08.008>
- Butts RA, Floate KD, David M et al (2003) Influence of intercropping canola or pea with barley on assemblages of ground beetles (Coleoptera: Carabidae). *Environ Entomol* 32:535–541. <https://doi.org/10.1603/0046-225X-32.3.535>
- Cadoux S, Sauzet G, Valantin-Morison M et al (2015) Intercropping frost-sensitive legume crops with winter oilseed rape reduces weed competition, insect damage, and improves nitrogen use efficiency. *OCL* 22:D302. <https://doi.org/10.1051/ocl/2015014>
- Cahenzli F, Sigsgaard L, Daniel C et al (2019) Perennial flower strips for pest control in organic apple orchards - a pan-European study. *Agric Ecosyst Environ* 278:43–53. <https://doi.org/10.1016/j.agee.2019.03.011>
- Campbell A, Wilby A, Sutton P, Wäckers F (2017a) Getting more power from your flowers: multi-functional flower strips enhance pollinators and pest control agents in Apple orchards. *Insects* 8:101. <https://doi.org/10.3390/insects8030101>
- Campbell AJ, Wilby A, Sutton P, Wäckers FL (2017b) Do sown flower strips boost wild pollinator abundance and pollination services in a spring-flowering crop? A case study from UK cider apple orchards. *Agric Ecosyst Environ* 239:20–29. <https://doi.org/10.1016/j.agee.2017.01.005>
- Campbell J, Miller D, Martin J (2016) Switchgrass (*Panicum virgatum*) intercropping within managed loblolly pine (*Pinus taeda*) does not affect wild bee communities. *Insects* 7:62. <https://doi.org/10.3390/insects7040062>
- Campera M, Balestri M, Manson S et al (2021) Shade trees and agrochemical use affect butterfly assemblages in coffee home gardens. *Agric Ecosyst Environ* 319:107547. <https://doi.org/10.1016/j.agee.2021.107547>
- Carvalho MG, Bortolotto OC, Ventura MU (2017) Aromatic plants affect the selection of host tomato plants by *B. EMISIA TABACI* biotype B. *Entomol Exp Appl* 162:86–92. <https://doi.org/10.1111/eea.12534>
- Cavanagh AF, Adler LS, Hazzard RV (2010) Buttercup squash provides a marketable alternative to Blue Hubbard as a trap crop for control of striped cucumber beetles (Coleoptera: Chrysomelidae). *Environ Entomol* 39:1953–1960. <https://doi.org/10.1603/EN10056>
- Chabi-Olaye A, Nolte C, Schulthess F, Borgemeister C (2005a) Abundance, dispersion and parasitism of the stem borer *Busseola fusca* (Lepidoptera: Noctuidae) in maize in the humid forest zone of southern Cameroon. *Bull Entomol Res* 95:169–177. <https://doi.org/10.1079/BER2004347>
- Chabi-Olaye A, Nolte C, Schulthess F, Borgemeister C (2005b) Relationships of intercropped maize, stem borer damage to maize yield and land-use efficiency in the humid forest of Cameroon. *Bull Entomol Res* 95:417–427. <https://doi.org/10.1079/BER2005373>
- Chen B, Wang J, Zhang L et al (2011a) Effect of intercropping pepper with sugarcane on populations of *Liriomyza huidobrensis* (Diptera: Agromyzidae) and its parasitoids. *Crop Prot* 30:253–258. <https://doi.org/10.1016/j.cropro.2010.11.005>
- Chen L-L, You M-S, Chen S-B (2011b) Effects of cover crops on spider communities in tea plantations. *Biol Control* 59:326–335. <https://doi.org/10.1016/j.biocontrol.2011.09.007>
- Christmann S, Bencharhi Y, Anougmar S et al (2021) Farming with alternative pollinators benefits pollinators, natural enemies, and yields, and offers transformative change to agriculture. *Sci Rep* 11:18206. <https://doi.org/10.1038/s41598-021-97695-5>
- Cline GR, Sedlacek JD, Hillman SL et al (2008) Organic management of cucumber beetles in watermelon and muskmelon production. *HortTechnology* 18:436–444
- Coll M, Bottrell DG (1994) Effects of nonhost plant on an insect herbivore in diverse habitats. *Ecology* 75:723–731. <https://doi.org/10.2307/1941730>
- Coll M, Bottrell DG (1995) Predator-prey association in mono- and dicultures: effect of maize and bean vegetation. *Agric Ecosyst Environ* 54:115–125
- Costello M (1995) Abundance, growth rate and parasitism of *Brevicoryne brassicae* and *Myzus persicae* (Homoptera: Aphididae) on broccoli grown in living mulches. *Agric Ecosyst Environ* 52:187–196. [https://doi.org/10.1016/0167-8809\(94\)00535-M](https://doi.org/10.1016/0167-8809(94)00535-M)
- Cotes B, Rämert B, Nilsson U (2018) A first approach to pest management strategies using trap crops in organic carrot fields. *Crop Prot* 112:141–148. <https://doi.org/10.1016/j.cropro.2018.05.025>
- Cuperus F, Ozinga WA, Bianchi FJJA et al (2023) Effects of field-level strip and mixed cropping on aerial arthropod and arable flora communities. *Agric Ecosyst Environ* 354:108568. <https://doi.org/10.1016/j.agee.2023.108568>
- Dainese M, Montecchiari S, Sitzia T et al (2017) High cover of hedgerows in the landscape supports multiple ecosystem services in Mediterranean cereal fields. *J Appl Ecol* 54:380–388. <https://doi.org/10.1111/1365-2664.12747>
- De La Fuente EB, Suárez SA, Lenardis AE, Poggio SL (2014) Intercropping sunflower and soybean in intensive farming systems: evaluating yield advantage and effect on weed and insect assemblages. *NJAS Wageningen J Life Sci* 70(1):47–52. <https://doi.org/10.1016/j.njas.2014.05.002>
- DeGooyer TA, Pedigo LP, Rice ME (1999) Effect of alfalfa-grass intercrops on insect populations. *Environ Entomol* 28:703–710. <https://doi.org/10.1093/ee/28.4.703>
- Delaney A, Dembele A, Nombri I et al (2020) Local-scale tree and shrub diversity improves pollination services to shea trees in

- tropical West African parklands. *J Appl Ecol* 57:1504–1513. <https://doi.org/10.1111/1365-2664.13640>
- Den Belder E, Valcheva RI, Guldemond JA (1999) Increased damage by western flower thrips *Frankliniella occidentalis* in chrysanthemum intercropped with subterranean clover. *Entomol Exp Appl* 91:275–285. <https://doi.org/10.1046/j.1570-7458.1999.00494.x>
- Depalo L, Burgio G, Von Fragstein P et al (2017) Impact of living mulch on arthropod fauna: analysis of pest and beneficial dynamics on organic cauliflower (*Brassica oleracea* L. var. botrytis) in different European scenarios. *Renew Agric Food Syst* 32:240–247. <https://doi.org/10.1017/S1742170516000156>
- Devi S, Ram P, Rolania K (2020) Effect of intercropping on the parasitoids, *Encarsia* spp. and *Trichogramma* spp. in cotton fields, India. *Egypt J Biol Pest Control* 30:71. <https://doi.org/10.1186/s41938-020-00275-4>
- Dhandapani S, Pakkirisamy M, Rajaraman R et al (2024) Floral interventions enhance flower visitor communities and pollination services in moringa plantations. *J Appl Ecol* 61:90–102. <https://doi.org/10.1111/1365-2664.14532>
- Dingha BN, Omaliko PC, Amoah BA et al (2021) Evaluation of cowpea (*Vigna unguiculata*) in an intercropping system as pollinator enhancer for increased crop yield. *Sustainability* 13:9612. <https://doi.org/10.3390/su13179612>
- Dissemond A, Hindorf H (1990) Influence of sorghum/maize/cowpea intercropping on the insect situation at Mbita/Kenya. *J Appl Entomol* 109:144–150. <https://doi.org/10.1111/j.1439-0418.1990.tb00030.x>
- Ditner N, Balmer O, Beck J et al (2013) Effects of experimentally planting non-crop flowers into cabbage fields on the abundance and diversity of predators. *Biodivers Conserv* 22:1049–1061. <https://doi.org/10.1007/s10531-013-0469-5>
- Ditzler L, Rossing WAH, Schulte RPO et al (2023) Prospects for increasing the resolution of crop diversity for agroecosystem service delivery in a Dutch arable system. *Agric Ecosyst Environ* 351:108472. <https://doi.org/10.1016/j.agee.2023.108472>
- Dixon PL, Coady JR, Larson DJ, Spaner D (2004) Undersowing rutabaga with white clover: impact on *Delia radicum* (Diptera: Anthomyiidae) and its natural enemies. *Can Entomol* 136:427–442. <https://doi.org/10.4039/n03-067>
- Djuideu CTL, Bisseleua HDB, Kekeunou S, Ambele FC (2021) Rehabilitation practices in cocoa agroforestry systems mitigate outbreaks of termites and support cocoa tree development and yield. *Agric Ecosyst Environ* 311:107324. <https://doi.org/10.1016/j.agee.2021.107324>
- Dogramaci M, Shrefler JW, Roberts BW et al (2004) Comparison of management strategies for squash bugs (Hemiptera: Coreidae) in watermelon. *J Econ Entomol*. <https://doi.org/10.1093/jee/97.6.1999>
- Dong Z, Xia M, Li C et al (2021) A comparison of flower and grass strips for augmentation of beneficial arthropods in apple orchards. *Front Sustain Food Syst* 5:697864. <https://doi.org/10.3389/fsufs.2021.697864>
- Echezona BC (2002) Corn-stalk lodging and borer damage as influenced by varying corn densities and planting geometry with soybean (*Glycine max.* L. Merrill). *Int Agrophys* 21
- Eduardo WI, Silva AC, Volpe HXL et al (2023) Push-pull and kill strategy for *Diaphorina citri* control in citrus orchards. *Entomol Exp Appl* 171:287–299. <https://doi.org/10.1111/eea.13273>
- English-Loeb G, Rhainds M, Martinson T, Ugine T (2003) Influence of flowering cover crops on *Anagrus* parasitoids (Hymenoptera: Mymaridae) and *Erythroneura* leafhoppers (Homoptera: Cicadellidae) in New York vineyards. *Agric Entomol* 5:173–181. <https://doi.org/10.1046/j.1461-9563.2003.00179.x>
- Esquivel IL, Brewer MJ, Coulson RN (2020) Field edge and field-to-field ecotone-type influences on two cotton herbivores: cotton fleahopper, *Pseudatomoscelis seriatus* (Hemiptera: Miridae), and Verde plant bug, *Creontiades signatus*. *J Econ Entomol* 113:2213–2222. <https://doi.org/10.1093/jee/toaa137>
- Fathi SAA (2022) The role of intercrops of eggplant and cowpea on the control of *Leucinodes orbonalis* guenee (Lepidoptera: Crambidae). *Biocontrol* 67:307–317. <https://doi.org/10.1007/s10526-022-10140-y>
- Fattah AS, Arrahman A, et al (2023) Effect of the number of rows and cultivars of soybeans on damage intensity of pest and predator populations in corn-soybean intercropping, South Sulawesi Indonesia. *Legume Res*. <https://doi.org/10.18805/LRF-742>
- Feltham H, Park K, Minderman J, Goulson D (2015) Experimental evidence that wildflower strips increase pollinator visits to crops. *Ecol Evol* 5:3523–3530. <https://doi.org/10.1002/ece3.1444>
- Fernandes FS, Ramalho FS, Nascimento A et al (2012) Within-plant distribution of cotton aphid (Hemiptera: Aphididae), in cotton with colored fibers and cotton-fennel intercropping system. *Ann Entomol Soc Am* 105:599–607. <https://doi.org/10.1603/AN12018>
- Ferrante M, Schulze M, Westphal C (2024) Hedgerows can increase predation rates in wheat fields in homogeneous agricultural landscapes. *J Environ Manage* 349:119498. <https://doi.org/10.1016/j.jenvman.2023.119498>
- Flausino BF, Machado CFM, Silva JHC et al (2022) Intercropping maize with brachiaria can be a double-edged sword strategy. *Pest Manage Sci* 78:5243–5250. <https://doi.org/10.1002/ps.7143>
- Forehand LM, Orr DB, Linker HM (2006) Evaluation of a commercially available beneficial insect habitat for management of Lepidoptera pests. *J Econ Entomol*. <https://doi.org/10.1093/jee/99.3.641>
- Foti MC, Peri E, Wajnberg E et al (2019) Contrasting olfactory responses of two egg parasitoids to buckwheat floral scent are reflected in field parasitism rates. *J Pest Sci* 92:747–756. <https://doi.org/10.1007/s10340-018-1045-2>
- Frank DL, Liburd OE (2005) Effects of living and synthetic mulch on the population dynamics of whiteflies and aphids, their associated natural enemies, and insect-transmitted plant diseases in zucchini. *Environ Entomol* 34:857–865. <https://doi.org/10.1603/0046-225X-34.4.857>
- Fusser MS, Holland JM, Jeanneret P et al (2018) Interactive effects of local and landscape factors on farmland carabids. *Agric Entomol* 20:549–557. <https://doi.org/10.1111/afe.12288>
- Ganser D, Mayr B, Albrecht M, Knop E (2018) Wildflower strips enhance pollination in adjacent strawberry crops at the small scale. *Ecol Evol* 8:11775–11784. <https://doi.org/10.1002/ece3.4631>
- Gao H, Jia S, Liu Y et al (2024a) Influence of wheat-walnut intercropping on the *Sitobion avenae* and its predatory natural enemies. *J Asia-Pac Entomol* 27:102188. <https://doi.org/10.1016/j.aspen.2023.102188>
- Gao J, Tang J, Zhang S, Zhang C (2024b) Intercropped *Flemingia macrophylla* successfully traps tea aphid (*Toxoptera aurantia*) and alters associated networks to enhance tea quality. *Pest Manage Sci* 80:1474–1483. <https://doi.org/10.1002/ps.7879>
- George D, Port G, Collier R (2019) Living on the edge: using and improving trap crops for flea beetle management in small-scale cropping systems. *Insects* 10:286. <https://doi.org/10.3390/insects10090286>
- Gianoli E, Ramos I, Alfaro-Tapia A et al (2006) Benefits of a maize-bean-weeds mixed cropping system in Urubamba Valley, Peruvian Andes. *Int J Pest Manage* 52:283–289. <https://doi.org/10.1080/09670870600796722>
- Gold CS, Altieri MA, Bellotti AC (1989) The effects of intercropping and mixed varieties of predators and parasitoids of cassava whiteflies (Hemiptera: Aleyrodidae) in Colombia. *Bull Entomol Res* 79:115–121. <https://doi.org/10.1017/S0007485300018629>

- González-Rodríguez ÁL (2003) Dynamics of diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae), in cabbage under intercropping, biological control and Bt-based sprays. *JOURNAL OF AGRICULTURE-UNIVERSITY OF PUERTO RICO* 87:31–50
- Gruss I, Twardowski JP, Hurej M, Kozak M (2018) Effect of intercropping narrow-leafed lupin with spring triticale on the abundance and diversity of rove beetles. *Biotechnol Agron Soc Environ* 22:220–229
- Guenat S, Kaartinen R, Jonsson M (2019) Shade trees decrease pest abundances on brassica crops in Kenya. *Agroforest Syst* 93:641–652. <https://doi.org/10.1007/s10457-017-0159-5>
- Gurr GM, Lu Z, Zheng X et al (2016) Multi-country evidence that crop diversification promotes ecological intensification of agriculture. *Nat Plants* 2:16014. <https://doi.org/10.1038/nplants.2016.14>
- Hagler JR, Nieto DJ, Machtley SA, et al (2018) Dynamics of predation on *Lygus hesperus* (Hemiptera: Miridae) in Alfalfa trap-cropped organic strawberry. *J Insect Sci* 18. <https://doi.org/10.1093/jisesa/iey077>
- Hailu G, Niassy S, Zeyaur KR et al (2018) Maize-legume intercropping and push-pull for management of fall armyworm, stemborers, and *Striga* in Uganda. *Agron J* 110:2513–2522. <https://doi.org/10.2134/agronj2018.02.0110>
- Hansen LM, Lorentsen L, Boelt B (2008) How to reduce the incidence of black bean aphids (*Aphis fabae* Scop.) attacking organic growing field beans (*Vicia faba* L.) by growing partially resistant bean varieties and by intercropping field beans with cereals. *Acta Agric Scand Sect B* 58:359–364. <https://doi.org/10.1080/09064710701788844>
- HansPetersen HN, McSorley R, Liburd OE (2010) The impact of intercropping squash with non-crop vegetation borders on the above-ground arthropod community. *Fla Entomol* 93:590–608. <https://doi.org/10.1653/024.093.0418>
- Harelimana A, Le Goff G, Rukazambuga D, Hance T (2024) Coffee trees intercropped with common beans: an opportunity to regulate the aphid *Toxoptera aurantii* (Boyer de Fonscolombe) (Hemiptera: Aphididae) in coffee agroecosystems. *Arthropod-Plant Interact* 18:307–316. <https://doi.org/10.1007/s11829-023-10031-8>
- Haro-Barchin E, Scheper J, Ganuza C et al (2018) Landscape-scale forest cover increases the abundance of *Drosophila suzukii* and parasitoid wasps. *Basic Appl Ecol* 31:33–43. <https://doi.org/10.1016/j.baae.2018.07.003>
- Harterreiten-Souza ÉS, Togni PHB, Pires CSS, Sujii ER (2014) The role of integrating agroforestry and vegetable planting in structuring communities of herbivorous insects and their natural enemies in the Neotropical region. *Agroforest Syst* 88:205–219. <https://doi.org/10.1007/s10457-013-9666-1>
- Harvey CT, Eubanks MD (2004) Effect of habitat complexity on biological control by the red imported fire ant (Hymenoptera: Formicidae) in collards. *Biol Control* 29:348–358. <https://doi.org/10.1016/j.biocontrol.2003.08.006>
- Harvey CT, Eubanks MD (2005) Intraguild predation of parasitoids by *Solenopsis invicta*: a non-disruptive interaction. *Entomol Exp Appl* 114:127–135. <https://doi.org/10.1111/j.1570-7458.2005.00250.x>
- HassalP M, Hawthorne A, Maudsleyb M (1992) Effects of headland management on invertebrate communities in cereal fields. *Agric Ecosyst Environ* 40:155–178
- Hatt S, Lopes T, Boeraeve F et al (2017) Pest regulation and support of natural enemies in agriculture: experimental evidence of within field wildflower strips. *Ecol Eng* 98:240–245. <https://doi.org/10.1016/j.ecoleng.2016.10.080>
- Hatt S, Xu Q, Francis F, Chen J (2019) Intercropping oilseed rape with wheat and releasing *Harmonia axyridis* sex pheromone in Northern China failed to attract and support natural enemies of aphids. *Biotechnol Agron Soc Environ* 147–152. <https://doi.org/10.25518/1780-4507.17921>
- Hausammann A (1996) Strip-management in rape crop: is winter rape endangered by negative impacts of sown weed strips? *J Appl Entomol* 120:505–512. <https://doi.org/10.1111/j.1439-0418.1996.tb01643.x>
- Held DW, Gonsiska P, Potter DA (2003) Evaluating companion planting and non-host masking odors for protecting roses from the Japanese beetle (Coleoptera: Scarabaeidae). *J Econ Entomol*. <https://doi.org/10.1093/jee/96.1.81>
- Helenius J (1991) Insect Numbers and Pest Damage in Intercrops vs. Monocrops: Concepts and Evidence from a System of Faba Bean, Oats and *Rhopalosiphum padi* (Homoptera, Aphididae). *J Sustainable Agric Environ* 1:57–80. https://doi.org/10.1300/J064v01n03_06
- Hendges ARADA, Melo JWDS, Guimaraes MDA, Rabelo JDS (2018) Intercropping kale with culinary herbs alters arthropod diversity and hinders population growth in aphids. *HortScience* 53:44–48. <https://doi.org/10.21273/HORTSCI12010-17>
- Hinds J, Hooks CRR (2013) Population dynamics of arthropods in a sunn-hemp zucchini interplanting system. *Crop Prot* 53:6–12. <https://doi.org/10.1016/j.cropro.2013.06.003>
- Hinds J, Wang K-H, Hooks CRR (2016) Growth and yield of zucchini squash (*Cucurbita pepo* L.) as influenced by a sunn hemp living mulch. *Biol Agric Hort* 32:21–33. <https://doi.org/10.1080/01448765.2015.1017736>
- Hodgkiss D, Brown MJF, Fountain MT (2019) The effect of within-crop floral resources on pollination, aphid control and fruit quality in commercial strawberry. *Agric Ecosyst Environ* 275:112–122. <https://doi.org/10.1016/j.agee.2019.02.006>
- Hogg BN, Nelson EH, Mills NJ, Daane KM (2011) Floral resources enhance aphid suppression by a hoverfly. *Entomol Exp Appl* 141:138–144. <https://doi.org/10.1111/j.1570-7458.2011.01174.x>
- Holmes DM, Barrett GW (1997) Japanese beetle (*Popillia japonica*) dispersal behavior in intercropped vs. monoculture soybean agroecosystems. *Am Midl Nat* 137:312. <https://doi.org/10.2307/2426850>
- Hood ASC, Advento AD, Stone J et al (2020) Removing understory vegetation in oil palm agroforestry reduces ground-foraging ant abundance but not species richness. *Basic Appl Ecol* 48:26–36. <https://doi.org/10.1016/j.baae.2020.07.002>
- Hooks CR, Johnson MW (2004) Using undersown clovers as living mulches: effects on yields, lepidopterous pest infestations, and spider densities in a Hawaiian broccoli agroecosystem. *Int J Pest Manage* 50:115–120. <https://doi.org/10.1080/09670870410001663462>
- Hooks CRR, Johnson MW (2002) Lepidopteran pest populations and crop yields in row intercropped broccoli. *Agric for Entomol* 4:117–125. <https://doi.org/10.1046/j.1461-9563.2002.00129.x>
- Hooks CRR, Johnson MW (2006) Population densities of herbivorous lepidopterans in diverse cruciferous cropping habitats: effects of mixed cropping and using a living mulch. *Biocontrol* 51:485–506. <https://doi.org/10.1007/s10526-006-9009-5>
- Hooks CRR, Valenzuela HR, Defrank J (1998) Incidence of pests and arthropod natural enemies in zucchini grown with living mulches. *Agric Ecosyst Environ* 69:217–231. [https://doi.org/10.1016/S0167-8809\(98\)00110-8](https://doi.org/10.1016/S0167-8809(98)00110-8)
- Horgan FG, Ramal AF, Villegas JM et al (2017) Effects of bund crops and insecticide treatments on arthropod diversity and herbivore regulation in tropical rice fields. *J Appl Entomol* 141:587–599. <https://doi.org/10.1111/jen.12383>
- Howard C, Fountain MT, Brittain C et al (2024) Perennial flower margins reduce orchard fruit damage by rosy apple aphid, *Dysaphis plantaginea* (Homoptera: Aphididae). *J Appl Ecol* 61:821–835. <https://doi.org/10.1111/1365-2664.14598>

- Hüber C, Zettl F, Hartung J, Müller-Lindenlauf M (2022) The impact of maize-bean intercropping on insect biodiversity. *Basic Appl Ecol* 61:1–9. <https://doi.org/10.1016/j.baae.2022.03.005>
- Hudson TB, Alford AM, Bilbo TR et al (2023) Living mulches reduce natural enemies when combined with frequent pesticide applications. *Agric Ecosyst Environ* 357:108680. <https://doi.org/10.1016/j.agee.2023.108680>
- Hughes J (2000) Management of Diamondback Moth (Lepidoptera: Plutellidae) in cabbage using collard as a trap crop. *HortScience* 35:875–879
- Hummel JD, Dossdall LM, Clayton GW et al (2012) Ground beetle (Coleoptera: Carabidae) diversity, activity density, and community structure in a diversified agroecosystem. *Environ Entomol* 41:72–80. <https://doi.org/10.1603/EN11072>
- Hummel JD, Dossdall LM, Clayton GW et al (2010) Responses of the parasitoids of *Delia radicum* (Diptera: Anthomyiidae) to the vegetational diversity of intercrops. *Biol Control* 55:151–158. <https://doi.org/10.1016/j.biocontrol.2010.08.004>
- Hunt LG, Dively G, Hooks CRR (2021) Flowering *Chamaecrista fasciculata* borders enhance natural enemy populations and improve grain quality in field corn. *Agric Ecosyst Environ* 306:107193. <https://doi.org/10.1016/j.agee.2020.107193>
- Hurej M, Twardowski JP (2006) The influence of yellow lupin intercropped with spring triticale on predatory carabid beetles (Coleoptera: Carabidae). *Eur J Entomol* 103:259–261. <https://doi.org/10.14411/eje.2006.031>
- Hurej M, Twardowski JP, Kozak M (2013) Weevil (Coleoptera: Curculionidae) assemblages in the fields of narrow-leaved lupin sown as pure stand and intercropped with spring triticale. *Zemdirbyste-Agriculture* 100:393–400. <https://doi.org/10.13080/z-a.2013.100.050>
- Hussain RI, Walcher R, Vogel N et al (2023) Effectiveness of flowers strips on insect's restoration in intensive grassland. *Agric Ecosyst Environ* 348:108436. <https://doi.org/10.1016/j.agee.2023.108436>
- Hyder M, Li Y, Raza MF et al (2023) Enhancing *Coccinella* beetle biological pest control via a floral approach in cucumber greenhouse. *Life* 13:2080. <https://doi.org/10.3390/life13102080>
- Imbert C, Papaix J, Husson L et al (2020a) Estimating population dynamics parameters of cabbage pests in temperate mixed apple tree-cabbage plots compared to control vegetable plots. *Crop Prot* 129:105037. <https://doi.org/10.1016/j.cropro.2019.105037>
- Imbert C, Papaix J, Husson L et al (2020b) Pests, but not predators, increase in mixed fruit tree-vegetable plots compared to control vegetable plots in a Mediterranean climate. *Agroforest Syst* 94:627–638. <https://doi.org/10.1007/s10457-019-00430-3>
- Inclán DJ, Dainese M, Cerretti P et al (2016) Spillover of tachinids and hoverflies from different field margins. *Basic Appl Ecol* 17:33–42. <https://doi.org/10.1016/j.baae.2015.08.005>
- Irvin NA, Scarratt SL, Wratten SD et al (2006) The effects of floral understoreys on parasitism of leafrollers (Lepidoptera: Tortricidae) on apples in New Zealand. *Agric for Entomol* 8:25–34. <https://doi.org/10.1111/j.1461-9555.2006.00285.x>
- Isbell F, Adler PR, Eisenhauer N et al (2017) Benefits of increasing plant diversity in sustainable agroecosystems. *J Ecol* 105:871–879. <https://doi.org/10.1111/1365-2745.12789>
- Jackson MD, Sisson VA (1998) Potential of *Nicotiana kawakamii* (Solanaceae) as a trap crop for protecting flue-cured tobacco from damage by *Heliothis virescens* (Lepidoptera: Noctuidae) larvae. *J Econ Entomol* 91:759–766. <https://doi.org/10.1093/jee/91.3.759>
- Jahnová Z, Knapp M, Boháč J, Tulachová M (2016) The role of various meadow margin types in shaping carabid and staphylinid beetle assemblages (Coleoptera: Carabidae, Staphylinidae) in meadow dominated landscapes. *J Insect Conserv* 20:59–69. <https://doi.org/10.1007/s10841-015-9839-5>
- Järvinen A, Himanen SJ, Raiskio S, Hyvönen T (2022) Intercropping of insect-pollinated crops supports a characteristic pollinator assemblage. *Agric Ecosyst Environ* 332:107930. <https://doi.org/10.1016/j.agee.2022.107930>
- Järvinen A, Hyvönen T, Raiskio S, Himanen SJ (2023) Intercropping shifts the balance between generalist arthropod predators and oilseed pests towards natural pest control. *Agric Ecosyst Environ* 348:108415. <https://doi.org/10.1016/j.agee.2023.108415>
- Ji X, Wang J, Dainese M et al (2022) Ground cover vegetation promotes biological control and yield in pear orchards. *J Appl Entomol* 146:262–271. <https://doi.org/10.1111/jen.12965>
- Ju Q, Ouyang F, Gu S et al (2019) Strip intercropping peanut with maize for peanut aphid biological control and yield enhancement. *Agric Ecosyst Environ* 286:106682. <https://doi.org/10.1016/j.agee.2019.106682>
- Juric I, Salzburger W, Luka H, Balmer O (2015) Molecular markers for *Diadegma* (Hymenoptera: Ichneumonidae) species distinction and their use to study the effects of companion plants on biocontrol of the diamondback moth. *Biocontrol* 60:179–187. <https://doi.org/10.1007/s10526-014-9637-0>
- Juventia SD, Rossing WAH, Ditzler L, Van Apeldoorn DF (2021) Spatial and genetic crop diversity support ecosystem service delivery: a case of yield and biocontrol in Dutch organic cabbage production. *Field Crops Res* 261:108015. <https://doi.org/10.1016/j.fcr.2020.108015>
- Kabi S, Karungi J, Sigsgaard L, Ssebuliba JM (2016) *Dysmicoccus brevipes* (Cockerell) occurrence and infestation behaviour as influenced by farm type, cropping systems and soil management practices. *Agric Ecosyst Environ* 222:23–29. <https://doi.org/10.1016/j.agee.2016.01.040>
- Kahl HM, Leslie AW, Hooks CRR (2019) Effects of Red Clover Living Mulch on Arthropod Herbivores and Natural Enemies, and Cucumber Yield. *Ann Entomol Soc Am* 112:356–364. <https://doi.org/10.1093/aesa/say036>
- Kahn BA, Rebek EJ, Brandenberger LP et al (2017) Companion planting with white yarrow or with feverfew for squash bug, *Anasa tristis* (Hemiptera: Coreidae), management on summer squash. *Pest Manage Sci* 73:1127–1133. <https://doi.org/10.1002/ps.4427>
- Karel AK (1991) Effects of plant populations and intercropping on the population patterns of bean flies on common beans. *Environ Entomol* 20:354–357. <https://doi.org/10.1093/ee/20.1.354>
- Karimzadeh J, Besharatnejad MH (2019) Ecological control of *Plutella xylostella* (Lepidoptera, Plutellidae) using trap cropping and Bt applications. *Arch Phytopathol Plant Prot* 52:1326–1347. <https://doi.org/10.1080/03235408.2019.1707930>
- Karssemeijer PN, Croijmans L, Gajendiran K et al (2024) Diverse cropping systems lead to higher larval mortality of the cabbage root fly (*Delia radicum*). *J Pest Sci* 97:337–353. <https://doi.org/10.1007/s10340-023-01629-1>
- Kebede Y, Baudron F, Bianchi F, Tittone P (2018a) Unpacking the push-pull system: assessing the contribution of companion crops along a gradient of landscape complexity. *Agric Ecosyst Environ* 268:115–123. <https://doi.org/10.1016/j.agee.2018.09.012>
- Kebede Y, Bianchi F, Baudron F et al (2018b) Implications of changes in land cover and landscape structure for the biocontrol potential of stemborers in Ethiopia. *Biol Control* 122:1–10. <https://doi.org/10.1016/j.biocontrol.2018.03.012>
- Keerthi MC, Suroshe SS, Doddachowdappa S et al (2023) Bio-intensive tactics for the management of invasive fall armyworm for organic maize production. *Plants* 12:685. <https://doi.org/10.3390/plants12030685>
- Kemmerling LR, McCarthy AC, Brown CS, Haddad NM (2023) Butterfly biodiversity increases with prairie strips and conservation

- management in row crop agriculture. *Insect Conserv Diversity* 16:828–837. <https://doi.org/10.1111/icad.12675>
- Kenny GJ, Chapman RB (1988) Effects of an intercrop on the insect pests, yield, and quality of cabbage. *N Z J Exp Agric* 16:67–72. <https://doi.org/10.1080/03015521.1988.10425616>
- Khokhar S, Rolania K (2022) Efficacy of different management modules against tomato fruit borer, *Helicoverpa armigera* (Hübner). *Int J Trop Insect Sci* 42:2731–2738. <https://doi.org/10.1007/s42690-022-00806-6>
- Kingazi N, Temu R, Sirima A, Jonsson M (2024) Tropical agroforestry supports insect pollinators and improves bean yield. *J Appl Ecol* 61:1067–1080. <https://doi.org/10.1111/1365-2664.14629>
- Kirsch F, Hass AL, Link W, Westphal C (2023) Intercrops as foraging habitats for bees: bees do not prefer sole legume crops over legume-cereal mixtures. *Agric Ecosyst Environ* 343:108268. <https://doi.org/10.1016/j.agee.2022.108268>
- Koji S, Khan ZR, Midega CAO (2007) Field boundaries of *Panicum maximum* as a reservoir for predators and a sink for *Chilo partellus*. *J Appl Entomol* 131:186–196. <https://doi.org/10.1111/j.1439-0418.2006.01131.x>
- Köneke A, Uesugi R, Herz A et al (2023) Effects of wheat undersowing and sweet alyssum intercropping on aphid and flea beetle infestation in white cabbage in Germany and Japan. *J Plant Dis Prot* 130:619–631. <https://doi.org/10.1007/s41348-023-00730-y>
- Kovács G, Kaasik R, Lof ME et al (2019) Effects of land use on infestation and parasitism rates of cabbage seed weevil in oilseed rape. *Pest Manage Sci* 75:658–666. <https://doi.org/10.1002/ps.5161>
- Kross SM, Martinico BL, Bourbour RP et al (2020) Effects of field and landscape scale habitat on insect and bird damage to sunflowers. *Front Sustain Food Syst* 4:40. <https://doi.org/10.3389/fsufs.2020.00040>
- Laffon L, Bischoff A, Gautier H et al (2022) Conservation biological control of codling moth (*Cydia pomonella*): effects of two aromatic plants, basil (*Ocimum basilicum*) and French marigolds (*Tagetes patula*). *Insects* 13:908. <https://doi.org/10.3390/insects13100908>
- Lai R, Hu H, Wu X et al (2019) Intercropping oilseed rape as a potential relay crop for enhancing the biological control of green peach aphids and aphid-transmitted virus diseases. *Entomol Exp Appl* 167:969–976. <https://doi.org/10.1111/eea.12850>
- Lai R, You M, Lotz LAP (Bert), Vasseur L (2011) Response of green peach aphids and other arthropods to garlic intercropped with tobacco. *Agron J* 103:856–863. <https://doi.org/10.2134/agronj2010.0404>
- Lale NES, Sastawa BM (2000) Evaluation of host plant resistance, sowing date modification and intercropping as methods for the control of *Mylabris* and *Coryna* species (Coleoptera: Meloidae) infesting pearl millet in the Nigerian Sudan savanna. *J Arid Environ* 46:263–280. <https://doi.org/10.1006/jare.2000.0690>
- Lamp WO (1991) Reduced *Empoasca fabae* (Homoptera: Cicadellidae) density in oat-alfalfa intercrop systems. *Environ Entomol* 20:118–126. <https://doi.org/10.1093/ee/20.1.118>
- Langer V (1996) Insect-crop interactions in a diversified cropping system: parasitism by *Aleochara bilineata* and *Trybliographa rapae* of the cabbage root fly, *Delia radicum*, on cabbage in the presence of white clover. *Entomol Exp Appl* 80:365–374. <https://doi.org/10.1111/j.1570-7458.1996.tb00949.x>
- Lee JC, Heimpel GE (2005) Impact of flowering buckwheat on Lepidopteran cabbage pests and their parasitoids at two spatial scales. *Biol Control* 34:290–301. <https://doi.org/10.1016/j.biocontrol.2005.06.002>
- Lee M, Campbell JW, Miller DA, Martin JA (2018) Insect community response to switchgrass intercropping and stand age of loblolly pine (*Pinus taeda*) plantations. *Agric for Entomol* 20:217–227. <https://doi.org/10.1111/afe.12247>
- Lemes PG (2014) Effect of intercropping on predation of *Oncideres ocularis* (Coleoptera: Cerambycidae) in Brazilian *Acacia mangium* plantations. *Rev Colomb Entomol* 40:34–39
- Lérault L, Clavel E, Villegas CM et al (2021) Providing alternative hosts and nectar to aphid parasitoids in a plum orchard to determine resource complementarity and distance range effect on biological control. *Agronomy* 12:77. <https://doi.org/10.3390/agronomy12010077>
- Leslie AW, Hamby KA, McCluen SR, Hooks CRR (2020) Evaluating a push-pull tactic for management of *Epilachna varivestis* Mulsant and enhancement of beneficial arthropods in *Phaseolus lunatus* L. *Ecol Eng* 147:105660. <https://doi.org/10.1016/j.ecoleng.2019.105660>
- Letourneau DK (1990) Mechanisms of predator accumulation in a mixed crop system. *Ecol Entomol* 15:63–69. <https://doi.org/10.1111/j.1365-2311.1990.tb00784.x>
- Letourneau DK (1995) Associational susceptibility: effects of cropping pattern and fertilizer on Malawian bean fly levels. *Ecol Appl* 5:823–829. <https://doi.org/10.2307/1941990>
- Li J, Zhou Y, Zhou B et al (2019) Habitat management as a safe and effective approach for improving yield and quality of tea (*Camellia sinensis*) leaves. *Sci Rep* 9:433. <https://doi.org/10.1038/s41598-018-36591-x>
- Li L, Chen F, Xing G (2022) Effects of fertilizer level and intercropping planting pattern with corn on the yield-related traits and insect community of soybean. *Agronomy* 12:3080. <https://doi.org/10.3390/agronomy12123080>
- Li S, Jaworski CC, Hatt S et al (2021a) Flower strips adjacent to greenhouses help reduce pest populations and insecticide applications inside organic commercial greenhouses. *J Pest Sci* 94:679–689. <https://doi.org/10.1007/s10340-020-01285-9>
- Li X, Lu X, Zhang Z et al (2021b) Intercropping rosemary (*Rosmarinus officinalis*) with sweet pepper (*Capsicum annuum*) reduces major pest population densities without impacting natural enemy populations. *Insects* 12:74. <https://doi.org/10.3390/insects12010074>
- Li Z, Yu J, Xu H et al (2023) Intercropping okra and castor bean reduces recruitment of Oriental fruit moth, *Grapholita molesta* (Lepidoptera: Tortricidae) in a pear orchard. *Insects* 14:885. <https://doi.org/10.3390/insects14110885>
- Liang K, Yang T, Zhang S et al (2016) Effects of intercropping rice and water spinach on net yields and pest control: an experiment in southern China. *Int J Agric Sustain* 14(4):448–465. <https://doi.org/10.1080/14735903.2016.1155391>
- Lin K, Lu Y, Wan P et al (2015) Simultaneous reduction in incidence of *Bemisia tabaci* (Hemiptera: Aleyrodidae) and *Sylepta derogata* (Lepidoptera: Pyralidae) using velvetleaf, *Abutilon theophrasti* as a trap crop. *J Pest Sci* 88:49–56. <https://doi.org/10.1007/s10340-014-0584-4>
- Litsinger JA, Hasse V, Barrion AT, Schmutterer H (1991) Response of *ostriaria furnacalis* (Guenée) (Lepidoptera: Pyralidae) to intercropping. *Environ Entomol* 20:988–1004. <https://doi.org/10.1093/ee/20.4.988>
- Liu J, Yan Y, Ali A et al (2017) Effects of wheat-maize intercropping on population dynamics of wheat aphids and their natural enemies. *Sustainability* 9:1390. <https://doi.org/10.3390/su9081390>
- Liu J-L, Ren W, Zhao W-Z, Li F-R (2018) Cropping systems alter the biodiversity of ground- and soil-dwelling herbivorous and predatory arthropods in a desert agroecosystem: implications for pest biocontrol. *Agric Ecosyst Environ* 266:109–121. <https://doi.org/10.1016/j.agee.2018.07.023>
- Lopez M, Liburd OE (2023) Effects of intercropping marigold, cowpea and an insecticidal soap on whiteflies and aphids in organic squash. *J Appl Entomol* 147:452–463. <https://doi.org/10.1111/jen.13141>

- Lu YH, Wu KM, Wyckhuys KAG, Guo YY (2009) Potential of mungbean, *Vigna radiata* as a trap crop for managing *Apolysus lucorum* (Hemiptera: Miridae) on Bt cotton. *Crop Prot* 28:77–81. <https://doi.org/10.1016/j.cropro.2008.08.018>
- Lubanga UK, Karungi J, Kyamanywa S, Ekbom B (2012) Assessing the potential of trap cropping in the management of different insect taxa on white cabbage. *Int J Trop Insect Sci* 32:218–223. <https://doi.org/10.1017/S1742758412000306>
- Ludwig SW, Kok LT (1998) Evaluation of trap crops to manage harlequin bugs, *Murgantia histrionica* (Hahn) (Hemiptera: Pentatomidae) on broccoli. *Crop Prot* 17:123–128. [https://doi.org/10.1016/S0261-2194\(97\)00107-5](https://doi.org/10.1016/S0261-2194(97)00107-5)
- Lun X, Jin M, Chen Z et al (2024) Flowering *Ocimum gratissimum* intercropped in tea plantations attracts and reduces *Apolysus lucorum* populations. *Pest Manage Sci* 80:4841–4852. <https://doi.org/10.1002/ps.8120>
- Luther GC, Valenzuela HR, Defrank J (1996) Impact of cruciferous trap crops on lepidopteran pests of cabbage in Hawaii. *Environ Entomol* 25:39–47. <https://doi.org/10.1093/ee/25.1.39>
- Maeda T, Hiraiwa MK, Ebata M et al (2023) *Brassica* plants promote *Apis mellifera* visitation to Japanese apricot in cold conditions. *Sci Hortic* 312:111844. <https://doi.org/10.1016/j.scienta.2023.111844>
- Manandhar R, Hooks CRR, Wright MG (2009) Influence of cover crop and intercrop systems on *Bemisia argentifolli* (Hemiptera: Aleyrodidae) infestation and associated squash silverleaf disorder in zucchini. *Environ Entomol* 38:442–449. <https://doi.org/10.1603/022.038.0218>
- Manandhar R, Wright MG (2015) Enhancing biological control of corn earworm, *Helicoverpa zea* and thrips through habitat management and inundative release of *Trichogramma pretiosum* in corn cropping systems. *Biol Control* 89:84–90. <https://doi.org/10.1016/j.biocontrol.2015.05.020>
- Manandhar R, Wright MG (2016) Effects of interplanting flowering plants on the biological control of corn earworm (Lepidoptera: Noctuidae) and thrips (Thysanoptera: Thripidae) in sweet corn. *J Econ Entomol* 109:113–119. <https://doi.org/10.1093/jee/tov306>
- Mansion-Vaquié A, Ferrante M, Cook SM et al (2017) Manipulating field margins to increase predation intensity in fields of winter wheat (*Triticum aestivum*). *J Appl Entomol* 141:600–611. <https://doi.org/10.1111/jen.12385>
- Mansion-Vaquié A, Wezel A, Ferrer A (2019) Wheat genotypic diversity and intercropping to control cereal aphids. *Agric Ecosyst Environ* 285:106604. <https://doi.org/10.1016/j.agee.2019.106604>
- Martin-Chave A, Béral C, Capowiez Y (2019a) Agroforestry has an impact on nocturnal predation by ground beetles and Opiliones in a temperate organic alley cropping system. *Biol Control* 129:128–135. <https://doi.org/10.1016/j.biocontrol.2018.10.009>
- Martin-Chave A, Béral C, Mazzia C, Capowiez Y (2019b) Agroforestry impacts the seasonal and diurnal activity of dominant predatory arthropods in organic vegetable crops. *Agroforest Syst* 93:2067–2083. <https://doi.org/10.1007/s10457-018-0309-4>
- Martini X, Pelz-Stelinski KS, Stelinski LL (2015) Absence of windbreaks and replanting citrus in solid sets increase density of Asian citrus psyllid populations. *Agric Ecosyst Environ* 212:168–174. <https://doi.org/10.1016/j.agee.2015.06.027>
- Mateos-Fierro Z, Fountain MT, Garratt MPD et al (2021) Active management of wildflower strips in commercial sweet cherry orchards enhances natural enemies and pest regulation services. *Agric Ecosyst Environ* 317:107485. <https://doi.org/10.1016/j.agee.2021.107485>
- Mateos-Fierro Z, Garratt MPD, Fountain MT et al (2023) The potential of wildflower strips to enhance pollination services in sweet cherry orchards grown under polytunnels. *J Appl Ecol* 60:1044–1055. <https://doi.org/10.1111/1365-2664.14394>
- Matteson PC (1982) The effects of intercropping with cereals and minimal permethrin applications on insect pests of cowpea and their natural enemies in Nigeria. *Trop Pest Manag* 28:372–380. <https://doi.org/10.1080/09670878209370743>
- McHugh NM, Moreby S, Lof ME et al (2020) The contribution of semi-natural habitats to biological control is dependent on sentinel prey type. *J Appl Ecol* 57:914–925. <https://doi.org/10.1111/1365-2664.13596>
- McKechnie IM, Thomsen CJM, Sargent RD (2017) Forested field edges support a greater diversity of wild pollinators in lowbush blueberry (*Vaccinium angustifolium*). *Agric Ecosyst Environ* 237:154–161. <https://doi.org/10.1016/j.agee.2016.12.005>
- Mei Z, De Groot GA, Kleijn D et al (2021) Flower availability drives effects of wildflower strips on ground-dwelling natural enemies and crop yield. *Agric Ecosyst Environ* 319:107570. <https://doi.org/10.1016/j.agee.2021.107570>
- Meyling NV, Navntoft S, Philipsen H et al (2013) Natural regulation of *Delia radicum* in organic cabbage production. *Agric Ecosyst Environ* 164:183–189. <https://doi.org/10.1016/j.agee.2012.09.019>
- Middleton EG, MacRae IV, Philips CR (2021) Floral plantings in large-scale commercial agroecosystems support both pollinators and arthropod predators. *Insects* 12:91. <https://doi.org/10.3390/insects12020091>
- Midega CAO, Bruce TJA, Pickett JA et al (2015) Climate-adapted companion cropping increases agricultural productivity in East Africa. *Field Crops Res* 180:118–125. <https://doi.org/10.1016/j.fcr.2015.05.022>
- Midega CAO, Khan ZR, Van Den Berg J et al (2008) Response of ground-dwelling arthropods to a ‘push–pull’ habitat management system: spiders as an indicator group. *J Appl Entomol* 132:248–254. <https://doi.org/10.1111/j.1439-0418.2007.01260.x>
- Midega CAO, Khan ZR, Van Den Berg J et al (2009) Non-target effects of the ‘push–pull’ habitat management strategy: Parasitoid activity and soil fauna abundance. *Crop Prot* 28:1045–1051. <https://doi.org/10.1016/j.cropro.2009.08.005>
- Midega CAO, Pittchar JO, Pickett JA et al (2018) A climate-adapted push-pull system effectively controls fall armyworm, *Spodoptera frugiperda* (J E Smith), in maize in East Africa. *Crop Prot* 105:10–15. <https://doi.org/10.1016/j.cropro.2017.11.003>
- Mollaei M, Fathi SAA, Nouri-Ganbalani G et al (2020) Effects of strip intercropping of canola with faba bean, field pea, garlic, or wheat on control of cabbage aphid and crop yield. *Plant Prot Sci* 57:59–65. <https://doi.org/10.17221/132/2019-PPS>
- Morandin LA, Kremen C (2013) Hedgerow restoration promotes pollinator populations and exports native bees to adjacent fields. *Ecol Appl* 23:829–839. <https://doi.org/10.1890/12-1051.1>
- Morandin LA, Long RF, Kremen C (2016) Pest control and pollination cost-benefit analysis of hedgerow restoration in a simplified agricultural landscape. *J Econ Entomol* 109:1020–1027. <https://doi.org/10.1093/jee/tow086>
- Morandin LA, Long RF, Kremen C (2014) Hedgerows enhance beneficial insects on adjacent tomato fields in an intensive agricultural landscape. *Agric Ecosyst Environ* 189:164–170. <https://doi.org/10.1016/j.agee.2014.03.030>
- Moreau TL, Warman PR, Hoyle J (2006) An evaluation of companion planting and botanical extracts as alternative pest controls for the Colorado potato beetle. *Biol Agric Hortic* 23:351–370. <https://doi.org/10.1080/01448765.2006.9755336>
- Moreno CR, Racelis AE (2015) Attraction, Repellence, and Predation: Role of Companion Plants in Regulating *Myzus persicae* (Sulzer) (Hemiptera: Aphidae) in Organic Kale Systems of South Texas. *Southwest Entomolog* 40:1–14. <https://doi.org/10.3958/059.040.0101>
- Mota L, Hevia V, Rad C et al (2022) Flower strips and remnant semi-natural vegetation have different impacts on pollination and

- productivity of sunflower crops. *J Appl Ecol* 59:2386–2397. <https://doi.org/10.1111/1365-2664.14241>
- Mulugeta T, Muluata B, Tekie H et al (2019) Phosphite alters the behavioral response of potato tuber moth (*Phthorimaea operculella*) to field-grown potato. *Pest Manage Sci* 75:616–621. <https://doi.org/10.1002/ps.5152>
- Muñoz AE, Amouroux P, Zaviezo T (2021a) Native flowering shrubs promote beneficial insects in avocado orchards. *Agric for Entomol* 23:463–472. <https://doi.org/10.1111/afe.12447>
- Muñoz AE, Plantegenest M, Amouroux P, Zaviezo T (2021b) Native flower strips increase visitation by non-bee insects to avocado flowers and promote yield. *Basic Appl Ecol* 56:369–378. <https://doi.org/10.1016/j.baae.2021.08.015>
- Mutiga SK, Gohole LS, Auma EO (2010) Effects of integrating companion cropping and nitrogen application on the performance and infestation of collards by *Brevicoryne brassicae*. *Entomol Exp Appl* 134:234–244. <https://doi.org/10.1111/j.1570-7458.2009.00952.x>
- Mwani CN, Nyaanga J, Cheruiyot EK, et al (2021) Intercropping and diverse field margin vegetation suppress bean aphid (Homoptera: Aphididae) infestation in dolichos (*Lablab purpureus* L.). *J Plant Prot Res* 61
- Mweke A, Akutse KS, Ulrichs C et al (2020) Integrated management of *Aphis craccivora* in cowpea using intercropping and entomopathogenic fungi under field conditions. *J Fungi* 6:60. <https://doi.org/10.3390/jof6020060>
- Nampala P, Adipala E, -Latigo MWO, et al (1999) Effect of cowpea monocultures and polycultures with sorghum and greengram on predatory arthropods. *Ann Appl Biol* 135:457–461. <https://doi.org/10.1111/j.1744-7348.1999.tb00874.x>
- Navasero MV, Calumpang SMF (2013) Influence of intercropping vegetable crops on the population of some Arthropods associated with eggplant (*Solanum melongena* L.) and damage potential of eggplant fruit and shoot borer (*Leucinodes orbonalis* Guenee) in the presence of radish (*Raphanus sativus* L.). *The Philippine Agricultural Scientist* 96
- Ndakidemi BJ, Mbega ER, Ndakidemi PA et al (2022) Plant-rich field margins influence natural predators of aphids more than intercropping in common bean. *Insects* 13:569. <https://doi.org/10.3390/insects13070569>
- Ndzana RA, Magro A, Bedoussac L et al (2014) Is there an associational resistance of winter pea–durum wheat intercrops towards *Acyrtosiphon pisum* Harris? *J Appl Entomol* 138:577–585. <https://doi.org/10.1111/jen.12119>
- Ngangambe MH, Mwatawala MW (2020) Effects of entomopathogenic fungi (EPFs) and cropping systems on parasitoids of fall armyworm (*Spodoptera frugiperda*) on maize in eastern central, Tanzania. *Biocontrol Sci Technol* 30:418–430. <https://doi.org/10.1080/09583157.2020.1726878>
- Ngeve J (2003) The cassava root mealybug (*Stictococcus vayssierei* Richard) (Homoptera: Stictococcidae): a threat to cassava production and utilization in Cameroon. *Int J Pest Manage* 49:327–333. <https://doi.org/10.1080/09670870310001603900>
- Nibouche S, Tibère R, Costet L (2019) *Erianthus arundinaceus* as a trap crop for the sugarcane stem borer *Chilo sacchariphagus*: field validation and disease risk assessment. *Crop Prot* 124:104877. <https://doi.org/10.1016/j.cropro.2019.104877>
- Nilsson L, Klatt BK, Smith HG (2021) Effects of flower-enriched ecological focus areas on functional diversity across scales. *Front Ecol Evol* 9:629124. <https://doi.org/10.3389/fevo.2021.629124>
- Nilsson U, Rännbäck L, Anderson P, Rämert B (2012) Herbivore response to habitat manipulation with floral resources: a study of the cabbage root fly. *J Appl Entomol* 136:481–489. <https://doi.org/10.1111/j.1439-0418.2011.01685.x>
- Ning C, Qu J, He L et al (2017) Improvement of yield, pest control and Si nutrition of rice by rice-water spinach intercropping. *Field Crops Res* 208:34–43. <https://doi.org/10.1016/j.fcr.2017.04.005>
- Noman MS, Maleque MA, Alam MZ et al (2013) Intercropping mustard with four spice crops suppresses mustard aphid abundance, and increases both crop yield and farm profitability in central Bangladesh. *Int J Pest Manage* 59:306–313. <https://doi.org/10.1080/09670874.2013.863408>
- Nyasani JO, Meyhöfer R, Subramanian S, -Poehling M (2012) Effect of intercrops on thrips species composition and population abundance on French beans in Kenya. *Entomol Exp Appl* 142:236–246. <https://doi.org/10.1111/j.1570-7458.2011.01217.x>
- Obanyi JN, Ogendo JO, Mulwa RMS et al (2023) Field margins and cropping system influence diversity and abundance of aphid natural enemies in *Lablab purpureus*. *J Appl Entomol* 147:439–451. <https://doi.org/10.1111/jen.13125>
- Ofuya TI (1991) Observations on insect infestation and damage in cowpea (*Vigna unguiculata*) intercropped with tomato (*Lycopersicon esculentum*) in a rain forest area of Nigeria. *Ex Agric* 27:407–412. <https://doi.org/10.1017/S0014479700019384>
- Ogol C (1999) Maize stem borer colonization, establishment and crop damage levels in a maize-leucaena agroforestry system in Kenya. *Agric Ecosyst Environ* 76:1–15. [https://doi.org/10.1016/S0167-8809\(99\)00077-8](https://doi.org/10.1016/S0167-8809(99)00077-8)
- Ogol CKPO, Spence JR, Keddie A (1998) Natural enemy abundance and activity in a maize-Leucaena agroforestry system in Kenya. *Environ Entomol* 27:1444–1451. <https://doi.org/10.1093/ee/27.6.1444>
- Oloo GW, Ogeda K (1990) The incidence of *Chilo partellus* (Swinh.) (Pyralidae) and the contribution of natural enemies to its mortality under intercropping system in Kenya. *Trop Pest Manag* 36:244–248. <https://doi.org/10.1080/09670879009371481>
- Page WW, Smith MC, Holt J, Kyetere D (1999) Intercrops, *Cicadulina* spp., and maize streak virus disease. *Ann Appl Biol* 135:385–393. <https://doi.org/10.1111/j.1744-7348.1999.tb00865.x>
- Pålsson J, Porcel M, Dekker T, Tasin M (2022) Attract, reward and disrupt: responses of pests and natural enemies to combinations of habitat manipulation and semiochemicals in organic apple. *J Pest Sci* 95:619–631. <https://doi.org/10.1007/s10340-021-01410-2>
- Pardon P, Mertens J, Reubens B et al (2020) *Juglans regia* (walnut) in temperate arable agroforestry systems: effects on soil characteristics, arthropod diversity and crop yield. *Renew Agric Food Syst* 35:533–549. <https://doi.org/10.1017/S1742170519000176>
- Parker JE, Crowder DW, Eigenbrode SD, Snyder WE (2016) Trap crop diversity enhances crop yield. *Agric Ecosyst Environ* 232:254–262. <https://doi.org/10.1016/j.agee.2016.08.011>
- Parsons CK, Dixon PL, Colbo M (2007) Relay cropping cauliflower with lettuce as a means to manage first-generation cabbage maggot (Diptera: Anthomyiidae) and minimize cauliflower yield loss. *J Econ Entomol*. <https://doi.org/10.1093/jee/100.3.838>
- Parthiban P, Chinniah C, Baskaran RKM, et al (2017) Impact of intercropping system to minimize the sucking pests incidence in groundnut (*Arachis hypogaea* Linnaeus). *Legume Res*. <https://doi.org/10.18805/LR-3770>
- Pascual-Villalobos MJ, Lacasa A, González A et al (2006) Effect of flowering plant strips on aphid and syrphid populations in lettuce. *Eur J Agron* 24:182–185. <https://doi.org/10.1016/j.eja.2005.07.003>
- Päts P, Ekbohm B, Skovgård H (1997) Influence of intercropping on the abundance, distribution and parasitism of *Chilo* spp. (Lepidoptera: Pyralidae) eggs. *Bull Entomol Res* 87:507–513. <https://doi.org/10.1017/S0007485300041377>
- Pease CG, Zalom FG (2010) Influence of non-crop plants on stink bug (Hemiptera: Pentatomidae) and natural enemy abundance in tomatoes. *J Appl Entomol* 134:626–636. <https://doi.org/10.1111/j.1439-0418.2009.01452.x>

- Pecheur E, Piqueray J, Monty A et al (2020) The influence of ecological infrastructures adjacent to crops on their carabid assemblages in intensive agroecosystems. *PeerJ* 8:e8094. <https://doi.org/10.7717/peerj.8094>
- Pekár S (1999) Effect of IPM practices and conventional spraying on spider population dynamics in an apple orchard. *Agric Ecosyst Environ* 73:155–166. [https://doi.org/10.1016/S0167-8809\(99\)00024-9](https://doi.org/10.1016/S0167-8809(99)00024-9)
- Peñalver-Cruz A, Alvarez D, Lavandero B (2020) Do hedgerows influence the natural biological control of woolly apple aphids in orchards? *J Pest Sci* 93:219–234. <https://doi.org/10.1007/s10340-019-01153-1>
- Pereira ALC, Taques TC, Valim JOS et al (2015) The management of bee communities by intercropping with flowering basil (*Ocimum basilicum*) enhances pollination and yield of bell pepper (*Capsicum annuum*). *J Insect Conserv* 19:479–486. <https://doi.org/10.1007/s10841-015-9768-3>
- Perfecto I, Vandermeer JH, Bautista GL et al (2004) Greater predation in shaded coffee farms: the role of resident neotropical birds. *Ecology* 85:2677–2681. <https://doi.org/10.1890/03-3145>
- Peters VE (2014) Intercropping with Shrub Species That Display a ‘Steady-State’ Flowering Phenology as a Strategy for Biodiversity Conservation in Tropical Agroecosystems. *PLoS ONE* 9:e90510. <https://doi.org/10.1371/journal.pone.0090510>
- Peters VE, Carroll CR, Cooper RJ et al (2013) The contribution of plant species with a steady-state flowering phenology to native bee conservation and bee pollination services. *Insect Conserv Diversity* 6:45–56. <https://doi.org/10.1111/j.1752-4598.2012.00189.x>
- Pfiffner L, Luka H, Schlatter C et al (2009) Impact of wildflower strips on biological control of cabbage lepidopterans. *Agric Ecosyst Environ* 129:310–314. <https://doi.org/10.1016/j.agee.2008.10.003>
- Pfister SC, Schäfer RB, Schirmel J, Entling MH (2015) Effects of hedgerows and riparian margins on aerial web-building spiders in cereal fields. *J Arachnol* 43:400–405. <https://doi.org/10.1636/0161-8202-43.3.400>
- Phillips BW, Gardiner MM (2016) Does local habitat management or large-scale landscape composition alter the biocontrol services provided to pumpkin agroecosystems? *Biol Control* 92:181–194
- Phoofolo MW, Giles KL, Elliott NC (2010) Effects of relay-intercropping sorghum with winter wheat, alfalfa, and cotton on lady beetle (Coleoptera: Coccinellidae) abundance and species composition. *Environ Entomol* 39:763–774. <https://doi.org/10.1603/EN09122>
- Piato K, Subía C, Pico J et al (2021) Organic farming practices and shade trees reduce pest infestations in *Robusta* coffee systems in Amazonia. *Life* 11:413. <https://doi.org/10.3390/life11050413>
- Piekarska-Boniecka H, Siatkowski M, Trzcíński P, Siatkowski I (2015) The impact of the vegetation of apple orchard edges on quantity and quality structure of predatory hoverflies (Diptera: Syrphidae) communities. *Turkish Journal of Entomology* 39. <https://doi.org/10.16970/te.92495>
- Pierre JF, Latournerie-Moreno L, Garruña R et al (2022) Effect of maize-legume intercropping on maize physio-agronomic parameters and beneficial insect abundance. *Sustainability* 14:12385. <https://doi.org/10.3390/su141912385>
- Pisani Gareau TL, Letourneau DK, Shennan C (2013) Relative densities of natural enemy and pest insects within California hedgerows. *Environ Entomol* 42:688–702. <https://doi.org/10.1603/EN12317>
- Pitan OOR, Filani CO (2014) Effect of intercropping cucumber *Cucumis sativus* (Cucurbitaceae) at different times with maize *Zea mays* (Poaceae) on the density of cucumber insect pests. *Int J Trop Insect Sci* 34:269–276. <https://doi.org/10.1017/S1742758414000435>
- Pitan OOR, Olatunde GO (2006) Effects of intercropping tomato (*Lycopersicon esculentum*) at different times with cowpea (*Vigna unguiculata*) or okra (*Abelmoschus esculentus*) on crop damage by major insect pests. *J Agric Sci* 144:361–368. <https://doi.org/10.1017/S0021859606006277>
- Pollier A, Tricault Y, Plantegenest M, Bischoff A (2019) Sowing of margin strips rich in floral resources improves herbivore control in adjacent crop fields. *Agric for Entomol* 21:119–129. <https://doi.org/10.1111/afe.12318>
- Ponti L, Altieri MA, Gutierrez AP (2007) Effects of crop diversification levels and fertilization regimes on abundance of *Brevicoryne brassicae* (L.) and its parasitization by *Diaeretiella rapae* (M’Intosh) in broccoli. *Agric for Entomol* 9:209–214. <https://doi.org/10.1111/j.1461-9563.2007.00330.x>
- Potapov AM, Dupérré N, Jochum M et al (2020) Functional losses in ground spider communities due to habitat structure degradation under tropical land-use change. *Ecology* 101:e02957. <https://doi.org/10.1002/ecy.2957>
- Prado SG, Collazo JA, Irwin RE (2018) Resurgence of specialized shade coffee cultivation: effects on pollination services and quality of coffee production. *Agric Ecosyst Environ* 265:567–575. <https://doi.org/10.1016/j.agee.2018.07.002>
- Prasifka JR, Schmidt N, Kohler KA et al (2006) Effects of living mulches on predator abundance and sentinel prey in a corn–soybean–forage rotation. *Environ Entomol* 35:1423–1431
- Puliga GA, Sprangers T, Huiting H, Dauber J (2024) Management practices influence biocontrol potential of generalist predators in maize cropping systems. *Entomol Exp Appl* 172:132–144. <https://doi.org/10.1111/eea.13395>
- Raderschall CA, Lundin O, Lindström SAM, Bommarco R (2022) Annual flower strips and honeybee hive supplementation differently affect arthropod guilds and ecosystem services in a mass-flowering crop. *Agric Ecosyst Environ* 326:107754. <https://doi.org/10.1016/j.agee.2021.107754>
- Ramvalho FS, Fernandes FS, Nascimento ARB et al (2012a) Feeding damage from cotton aphids, *Aphis gossypii* Glover (Hemiptera: Heteroptera: Aphididae), in cotton with colored fiber intercropped with fennel. *Ann Entomol Soc Am* 105:20–27. <https://doi.org/10.1603/AN11122>
- Ramvalho FS, Fernandes FS, Nascimento ARB et al (2012b) Assessment of fennel aphids (Hemiptera: Aphididae) and their predators in fennel intercropped with cotton with colored fibers. *J Econ Entomol* 105:113–119. <https://doi.org/10.1603/EC11219>
- Rämert B (1996) The influence of intercropping and mulches on the occurrence of polyphagous predators in carrot fields in relation to carrot fly (*Psila rosae* (F.)) (Dipt., Psilidae) damage. *J Appl Entomol* 120:39–46. <https://doi.org/10.1111/j.1439-0418.1996.tb01565.x>
- Rämerti B, Ekbohm B (1996) Intercropping as a management strategy against carrot rust fly (Diptera: Psilidae): a test of enemies and resource concentration hypotheses. *Environ Entomol* 25:1092–1100. <https://doi.org/10.1093/ee/25.5.1092>
- Ramsden MW, Menéndez R, Leather SR, Wäckers F (2015) Optimizing field margins for biocontrol services: the relative role of aphid abundance, annual floral resources, and overwinter habitat in enhancing aphid natural enemies. *Agric Ecosyst Environ* 199:94–104. <https://doi.org/10.1016/j.agee.2014.08.024>
- Ratnadass A, Zakari-Moussa O, Kadi-Kadi HA et al (2014) Potential of pigeon pea as a trap crop for control of fruit worm infestation and damage to okra. *Agric for Entomol* 16:426–433. <https://doi.org/10.1111/afe.12072>
- Razze JM, Liburd OE, Webb SE (2016) Intercropping buckwheat with squash to reduce insect pests and disease incidence and increase yield. *Agroecol Sustainable Food Syst* 40:863–891. <https://doi.org/10.1080/21683565.2016.1205541>

- Rea JH, Wratten SD, Sedcole R et al (2002) Trap cropping to manage green vegetable bug *Nezara viridula* (L.) (Heteroptera: Pentatomidae) in sweet corn in New Zealand. *Agric Entomol* 4:101–107. <https://doi.org/10.1046/j.1461-9563.2002.00130.x>
- Rezende MQ, Venzon M, Dos Santos PS et al (2021) Extrafloral nectary-bearing leguminous trees enhance pest control and increase fruit weight in associated coffee plants. *Agric Ecosyst Environ* 319:107538. <https://doi.org/10.1016/j.agee.2021.107538>
- Rhino B, Grechi I, Marliac G et al (2014) Corn as trap crop to control *Helicoverpa zea* in tomato fields: importance of phenological synchronization and choice of cultivar. *Int J Pest Manage* 60:73–81. <https://doi.org/10.1080/09670874.2014.900708>
- Ribeiro AL, Gontijo LM (2017) Alyssum flowers promote biological control of collard pests. *Biocontrol* 62:185–196. <https://doi.org/10.1007/s10526-016-9783-7>
- Richard R, Cahon T, Llandres AL et al (2020) Alley cropping agroforestry mediates carabid beetle distribution at a micro-habitat scale. *Agroforest Syst* 94:309–317. <https://doi.org/10.1007/s10457-019-00390-8>
- Risch S (1980) The population dynamics of several herbivorous beetles in a tropical agroecosystem: the effect of intercropping corn, beans and squash in Costa Rica. *J Appl Ecol* 17:593. <https://doi.org/10.2307/2402639>
- Rizk AM (2011) Effect of strip-management on the population of the Aphid, *Aphis craccivora* Koch and its associated predators by intercropping Faba bean, *Vicia faba* L. with coriander, *Coriandrum sativum* L. *Egypt J Biol Pest Control* 21
- Roda AL, Landis DA, Coggins ML (1997) Forage grasses elicit emigration of adult potato leafhopper (Homoptera: Cicadellidae) from alfalfa-grass mixtures. *Environ Entomol* 26:745–753. <https://doi.org/10.1093/ee/26.4.745>
- Rodenwald N, Sutcliffe LME, Leuschner C, Batáry P (2023) Weak evidence for biocontrol spillover from both flower strips and grassy field margins in conventional cereals. *Agric Ecosyst Environ* 355:108614. <https://doi.org/10.1016/j.agee.2023.108614>
- Ruhanen HM, Mofikoya AO, Vesterbacka A et al (2023) Trait-based cropping of brassicaceous plants: effects on ecosystem services and crop yield. *Biol Control* 187:105389. <https://doi.org/10.1016/j.biocontrol.2023.105389>
- Saeed Q, Zaka M, Saeed S, Bakhtawar M (2013) Lucerne as trap crop in wheat for Development of Predators Population Against Wheat Aphids (Aphididae: Homoptera). *Pak J Zool* 45
- Salamanca J, Pareja M, Rodriguez-Saona C et al (2015) Behavioral responses of adult lacewings, *Chrysoperla externa*, to a rose-aphid–coriander complex. *Biol Control* 80:103–112. <https://doi.org/10.1016/j.biocontrol.2014.10.003>
- Saldanha AV, Gontijo LM, Carvalho RMR et al (2019) Companion planting enhances pest suppression despite reducing parasitoid emergence. *Basic Appl Ecol* 41:45–55. <https://doi.org/10.1016/j.baae.2019.10.002>
- Santillano-Cázares J, Mendoza-Gómez A, Vázquez-Angulo JC et al (2019) The compromise of intercropping: biological pest control versus competition by crop species. *Southwest Entomol* 44:393. <https://doi.org/10.3958/059.044.0204>
- Santos LAO, Botelho Costa M, Lavigne C et al (2018) Influence of the margin vegetation on the conservation of aphid biological control in apple orchards. *J Insect Conserv* 22:465–474. <https://doi.org/10.1007/s10841-018-0074-8>
- Sardiñas HS, Kremen C (2015) Pollination services from field-scale agricultural diversification may be context-dependent. *Agric Ecosyst Environ* 207:17–25. <https://doi.org/10.1016/j.agee.2015.03.020>
- Sardiñas HS, Ponisio LC, Kremen C (2016) Hedgerow presence does not enhance indicators of nest-site habitat quality or nesting rates of ground-nesting bees. *Restor Ecol* 24:499–505. <https://doi.org/10.1111/rec.12338>
- Sastawa BM, Lawan M, Maina YT (2004) Management of insect pests of soybean: effects of sowing date and intercropping on damage and grain yield in the Nigerian Sudan savanna. *Crop Prot* 23:155–161. <https://doi.org/10.1016/j.cropro.2003.07.007>
- Scagliarini O, Ferrari R, Masetti A, Burgio G (2023) Trap cropping: an agroecological approach to management of flea beetles on sugar beet. *Crop Prot* 166:106174. <https://doi.org/10.1016/j.cropro.2022.106174>
- Schader C, Zaller JG, Köpke U (2005) Cotton-basil intercropping: effects on pests, yields and economical parameters in an organic field in Fayoum, Egypt. *Biol Agric Hortic* 23:59–72. <https://doi.org/10.1080/01448765.2005.9755308>
- Schellhorn NA, Sork VL (1997) The impact of weed diversity on insect population dynamics and crop yield in collards, *Brassica oleracea* (Brassicaceae). *Oecologia* 111:233–240. <https://doi.org/10.1007/s004420050230>
- Schmidt NP, O’Neal ME, Singer JW (2007) Alfalfa living mulch advances biological control of soybean aphid. *Environ Entomol*. <https://doi.org/10.1093/ee/36.2.416>
- Schoeny A, Lauvernay A, Lambion J et al (2019) The beauties and the bugs: a scenario for designing flower strips adapted to aphid management in melon crops. *Biol Control* 136:103986. <https://doi.org/10.1016/j.biocontrol.2019.05.005>
- Schulthess F, Chabi-Olaye A, Gounou S (2004) Multi-trophic level interactions in a cassava–maize mixed cropping system in the humid tropics of West Africa. *Bull Entomol Res* 94:261–272. <https://doi.org/10.1079/BER2004296>
- Seimandi-Corda G, Winkler J, Jenkins T et al (2024) Companion plants and straw mulch reduce cabbage stem flea beetle (*Psylliodes chrysocephala*) damage on oilseed rape. *Pest Manage Sci* 80:2333–2341. <https://doi.org/10.1002/ps.7641>
- Sekamatte BM, Ogenga-Latigo M, Russell-Smith A (2003) Effects of maize–legume intercrops on termite damage to maize, activity of predatory ants and maize yields in Uganda. *Crop Prot* 22:87–93. [https://doi.org/10.1016/S0261-2194\(02\)00115-1](https://doi.org/10.1016/S0261-2194(02)00115-1)
- Sekine T, Kanao K, Inawashiro S, Hori M (2021a) Insect pest management by intercropping with leafy daikon (*Raphanus sativus*) in cabbage fields. *Arthropod-Plant Interact* 15:669–681. <https://doi.org/10.1007/s11829-021-09848-y>
- Sekine T, Masuda T, Inawashiro S (2021b) Suppression effect of intercropping with barley on *Thrips tabaci* (Thysanoptera: Thripidae) in onion fields. *Appl Entomol Zool* 56:59–68. <https://doi.org/10.1007/s13355-020-00708-4>
- Serée L, Barbottin A, Chiron F et al (2023) Within-field floral resources have the potential to increase parasitism rates in winter oilseed rape pests more than resources at field margins. *Agric Ecosyst Environ* 344:108288. <https://doi.org/10.1016/j.agee.2022.108288>
- Sherawat SM, Butt A, Tahir HM (2012) Effect of *Brassica* strips on the population of aphids and arthropod predators in wheat ecosystem. *Pak J Zool* 44:173–179
- Shi X, Axmacher JC, Luo A et al (2023) Wild pollinator communities benefit from mixed cultivation of oilseed rape and milk vetch. *J Appl Entomol* 147:966–975. <https://doi.org/10.1111/jen.13192>
- Shrestha B, Finke DL, Piñero JC (2019) The ‘Botanical Triad’: The Presence of Insectary Plants Enhances Natural Enemy Abundance on Trap Crop Plants in an Organic Cabbage Agro-Ecosystem. *Insects* 10:181. <https://doi.org/10.3390/insects10060181>
- Sigsgaard L (1999) Effects of cowpea intersowing and insecticide application on *Helicoverpa armigera* Hubner (Lepidoptera: Noctuidae) and its natural enemies in pigeonpea intercropped with sorghum. *Int J Pest Manage* 45:61–67. <https://doi.org/10.1080/096708799228067>
- Silva EB, Franco JC, Vasconcelos T, Branco M (2010) Effect of ground cover vegetation on the abundance and diversity of beneficial

- arthropods in citrus orchards. *Bull Entomol Res* 100:489–499. <https://doi.org/10.1017/S0007485309990526>
- Singh A, Weisser WW, Hanna R et al (2017) Reduce pests, enhance production: benefits of intercropping at high densities for okra farmers in Cameroon. *Pest Manage Sci* 73:2017–2027. <https://doi.org/10.1002/ps.4636>
- Skelton LE, Barrett GW (2005) A comparison of conventional and alternative agroecosystems using alfalfa (*Medicago sativa*) and winter wheat (*Triticum aestivum*). *Renew Agric Food Syst* 20:38–47. <https://doi.org/10.1079/RAF200478>
- Skovgård H, Pääts P (1996) Effects of intercropping on maize stem borers and their natural enemies. *Bull Entomol Res* 86:599–607. <https://doi.org/10.1017/S0007485300039407>
- Smith HA, McSorley R, Izaguirre JAS (2001) Effect of intercropping common bean with poor hosts and nonhosts on numbers of immature whiteflies (Homoptera: Aleyrodidae) in the Salamá Valley, Guatemala. *Environ Entomol* 30:89–100. <https://doi.org/10.1603/0046-225X-30.1.89>
- Smith J, Potts S, Eggleton P (2008) The value of sown grass margins for enhancing soil macrofaunal biodiversity in arable systems. *Agric Ecosyst Environ* 127:119–125. <https://doi.org/10.1016/j.agee.2008.03.008>
- Smits N, Dupraz C, Dufour L (2012) Unexpected lack of influence of tree rows on the dynamics of wheat aphids and their natural enemies in a temperate agroforestry system. *Agroforest Syst* 85:153–164. <https://doi.org/10.1007/s10457-011-9473-5>
- Song B, Han Z (2020) Assessment of the biocontrol effects of three aromatic plants on multiple trophic levels of the arthropod community in an agroforestry ecosystem. *Ecol Entomol* 45:831–839. <https://doi.org/10.1111/een.12858>
- Song B, Jiao H, Tang G, Yao Y (2014) Combining repellent and attractive aromatic plants to enhance biological control of three tortricid species (Lepidoptera: Tortricidae) in an apple orchard. *Fla Entomol* 97:1679–1689. <https://doi.org/10.1653/024.097.0442>
- Song B, Tang G, Sang X et al (2013) Intercropping with aromatic plants hindered the occurrence of *Aphis citricola* in an apple orchard system by shifting predator–prey abundances. *Biocontrol Sci Technol* 23:381–395. <https://doi.org/10.1080/09583157.2013.763904>
- Song BZ, Wu HY, Kong Y et al (2010) Effects of intercropping with aromatic plants on the diversity and structure of an arthropod community in a pear orchard. *Biocontrol* 55:741–751. <https://doi.org/10.1007/s10526-010-9301-2>
- Songa JM, Jiang N, Schulthess F, Omwega C (2007) The role of intercropping different cereal species in controlling lepidopteran stem borers on maize in Kenya. *J Appl Entomol* 131:40–49. <https://doi.org/10.1111/j.1439-0418.2006.01116.x>
- Souza IL, Tomazella VB, Santos AJN et al (2019) Parasitoids diversity in organic sweet pepper (*Capsicum annuum*) associated with basil (*Ocimum basilicum*) and marigold (*Tagetes erecta*). *Braz J Biol* 79:603–611. <https://doi.org/10.1590/1519-6984.185417>
- Srinivasa Rao M, Rama Rao CA, Srinivas K et al (2012) Intercropping for management of insect pests of castor, *Ricinus communis*, in the semi-arid tropics of India. *J Insect Sci* 12:1–10. <https://doi.org/10.1673/031.012.1401>
- Srinivasan K, Moorthy PNK (1991) Indian mustard as a trap crop for management of major lepidopterous pests on cabbage. *Trop Pest Manag* 37:26–32. <https://doi.org/10.1080/09670879109371532>
- Stahl JM, Wilson H, Straser RK et al (2021) Irrigated trap crops impact key hemipteran pests in organic pistachio orchard. *Arthropod-Plant Interact* 15:949–959. <https://doi.org/10.1007/s11829-021-09869-7>
- Stamps WT, McGraw RL, Godsey L, Woods TL (2009a) The ecology and economics of insect pest management in nut tree alley cropping systems in the Midwestern United States. *Agric Ecosyst Environ* 131:4–8. <https://doi.org/10.1016/j.agee.2008.06.012>
- Stamps WT, Nelson EA, Linit MJ (2009b) Survey of diversity and abundance of ground-dwelling arthropods in a black walnut-forage alley-cropped system in the mid-western United States. *J Kans Entomol Soc* 82:46–62. <https://doi.org/10.2317/JKES705.01.1>
- Stathakis T, Economou L, Barda M et al (2023) Potential of hedgerows with aromatic plants as reservoirs of natural enemies of pests in orange orchards. *Insects* 14:391. <https://doi.org/10.3390/insects14040391>
- Staton T, Walters RJ, Smith J et al (2021) Evaluating a trait-based approach to compare natural enemy and pest communities in agroforestry vs. arable systems. *Ecol Appl* 31:e02294. <https://doi.org/10.1002/eap.2294>
- Staudacher K, Schallhart N, Thalinger B et al (2013) Plant diversity affects behavior of generalist root herbivores, reduces crop damage, and enhances crop yield. *Ecol Appl* 23:1135–1145. <https://doi.org/10.1890/13-0018.1>
- Stephens MJ, France CM, Wratten SD, Frampton C (1998) Enhancing biological control of leafrollers (Lepidoptera: Tortricidae) by sowing buckwheat (*Fagopyrum esculentum*) in an orchard. *Biocontrol Sci Technol* 8:547–558. <https://doi.org/10.1080/09583159830063>
- Straub CS, Faselt JA, Keyser ES, Traugott M (2020) Host plant resistance promotes a secondary pest population. *Ecosphere* 11:e03073. <https://doi.org/10.1002/ecs2.3073>
- Straub CS, Simasek NP, Dohm R et al (2014) Plant diversity increases herbivore movement and vulnerability to predation. *Basic Appl Ecol* 15:50–58. <https://doi.org/10.1016/j.baae.2013.12.004>
- Sujayanand GK, Chandra A, Pandey S, Bhatt S (2020) Seasonal Abundance of Spotted Pod Borer, *Maruca vitrata* Fabricius in Early Pigeonpea [*Cajanus cajan* (L.) Millsp.] and its Management through Farmscaping in Uttar Pradesh. *Legume Res*. <https://doi.org/10.18805/LR-4329>
- Sujayanand GK, Sharma RK, Shankarganesh K et al (2015) Crop diversification for sustainable insect pest management in eggplant (Solanales: Solanaceae). *Fla Entomol* 98:305–314. <https://doi.org/10.1653/024.098.0149>
- Sujayanand GK, Sharma RK, Shankarganesh K (2016) Impact of intercropping and border crops on pest incidence in okra. *Ind Jour of Hort* 73:219. <https://doi.org/10.5958/0974-0112.2016.00051.7>
- Sulvai F, Chaúque BJM, Macuvele DLP (2016) Intercropping of lettuce and onion controls caterpillar thread, *Agrotis ipsilon* major insect pest of lettuce. *Chem Biol Technol Agric* 3:28. <https://doi.org/10.1186/s40538-016-0079-z>
- Sutter L, Albrecht M, Jeanneret P (2018a) Landscape greening and local creation of wildflower strips and hedgerows promote multiple ecosystem services. *J Appl Ecol* 55:612–620. <https://doi.org/10.1111/1365-2664.12977>
- Sutter L, Amato M, Jeanneret P, Albrecht M (2018b) Overwintering of pollen beetles and their predators in oilseed rape and semi-natural habitats. *Agric Ecosyst Environ* 265:275–281. <https://doi.org/10.1016/j.agee.2018.06.030>
- Szendrei Z (2012) The impact of plant associations on *Macrostelus quadrilineatus* management in carrots. *Entomol Exp Appl* 143:191–198. <https://doi.org/10.1111/j.1570-7458.2012.01243.x>
- Tajmiri P, Fathi SAA, Golizadeh A, Nouri-Ganbalani G (2017a) Strip-intercropping canola with annual alfalfa improves biological control of *Plutella xylostella* (L.) and crop yield. *Int J Trop Insect Sci* 37:208–216. <https://doi.org/10.1017/S1742758417000145>

- Tajmiri P, Fathi SAA, Golizadeh A, Nouri-Ganbalani G (2017b) Effect of strip-intercropping potato and annual alfalfa on populations of *Leptinotarsa decemlineata* Say and its predators. *Int J Pest Manage* 63:273–279. <https://doi.org/10.1080/09670874.2016.1256513>
- Tang GB, Song BZ, Zhao LL et al (2013) Repellent and attractive effects of herbs on insects in pear orchards intercropped with aromatic plants. *Agroforest Syst* 87:273–285. <https://doi.org/10.1007/s10457-012-9544-2>
- Tanyi CB, Ngosong C, Ntonifor NN (2018) Effects of climate variability on insect pests of cabbage: adapting alternative planting dates and cropping pattern as control measures. *Chem Biol Technol Agric* 5:25. <https://doi.org/10.1186/s40538-018-0140-1>
- Theunissen J, Booij C, Lotz L (1995) Effects of intercropping white cabbage with clovers on pest infestation and yield. *Entomol Exp Appl* 74:7–16
- Thevathasan NV, Gordon AM (2004) Ecology of tree intercropping systems in the North temperate region: experiences from southern Ontario, Canada. *Agroforest Syst* 61:257–268
- Tillman P, Khrimian A, Cottrell T et al (2015) Trap cropping systems and a physical barrier for suppression of stink bugs (Hemiptera: Pentatomidae) in cotton. *J Econ Entomol* 108:2324–2334
- Tillman PG (2006a) Sorghum as a Trap Crop for *Nezara viridula* L. (Heteroptera: Pentatomidae) in Cotton in the Southern United States. *Environ Entomol* 35:771–783. <https://doi.org/10.1603/0046-225X-35.3.771>
- Tillman PG (2006b) Tobacco as a trap crop for *Heliothis virescens* (F.) (Lepidoptera: Noctuidae) in cotton. *J Entomol Sci* 41:305–320
- Tillman PG, Mullinix BG (2004) Grain sorghum as a trap crop for corn earworm (Lepidoptera: Noctuidae) in cotton. *Environ Entomol* 33:1371–1380. <https://doi.org/10.1603/0046-225X-33.5.1371>
- Tingey WM, Lamont WJ (1988) Insect abundance in field beans altered by intercropping. *Bull Entomol Res* 78:527–535. <https://doi.org/10.1017/S0007485300013274>
- Tiwari S, Saville DJ, Sharma S et al (2020a) Evaluation of potential trap plant species for the wheat bug *Nysius huttoni* (Hemiptera: Lygaeidae) in forage brassicas. *Agric for Entomol* 22:263–273. <https://doi.org/10.1111/afe.12379>
- Tiwari S, Sharma S, Wratten SD (2020b) Flowering alyssum (*Lobularia maritima*) promote arthropod diversity and biological control of *Myzus persicae*. *J Asia-Pac Entomol* 23:634–640. <https://doi.org/10.1016/j.aspen.2020.05.002>
- Toennisson TA, Klein JT, Burrack H (2019) Measuring the effect of non-crop flowering plants on natural enemies in organic tobacco. *Biol Control* 137:104023. <https://doi.org/10.1016/j.biocontrol.2019.104023>
- Togni PHB, Marouelli WA, Inoue-Nagata AK et al (2018) Integrated cultural practices for whitefly management in organic tomato. *J Appl Entomol* 142:998–1007. <https://doi.org/10.1111/jen.12558>
- Toivonen M, Huusela-Veistola E, Herzon I (2018) Perennial fallow strips support biological pest control in spring cereal in northern Europe. *Biol Control* 121:109–118. <https://doi.org/10.1016/j.biocontrol.2018.02.015>
- Tomazella VB, Jacques GC, Lira AC, Silveira LCP (2018) Visitation of social wasps in arabica coffee crop (*Coffea arabica* L.) intercropped with different tree species. *Sociobiology* 65:299. <https://doi.org/10.13102/sociobiology.v65i2.1397>
- Tonhasca A (1993) Effects of agroecosystem diversification on natural enemies of soybean herbivores. *Entomol Exp Appl* 69:83–90. <https://doi.org/10.1111/j.1570-7458.1993.tb01731.x>
- Török E, Zieger S, Rosenthal J et al (2021) Organic farming supports lower pest infestation, but fewer natural enemies than flower strips. *J Appl Ecol* 58:2277–2286. <https://doi.org/10.1111/1365-2664.13946>
- Tosti G, Falcinelli B, Guiducci M (2023) Lentil–cereal intercropping in a Mediterranean area: yield, pests and weeds. *Agron J* 115:2570–2578. <https://doi.org/10.1002/agj2.21413>
- Tougeron K, Ferrais L, Gardin P et al (2023) Flower strips increase the control of rosy apple aphids after parasitoid releases in an apple orchard. *Ann Appl Biol* 182:245–256. <https://doi.org/10.1111/aab.12816>
- Toukem NK, Dubois T, Mohamed SA et al (2023) The effect of annual flower strips on pollinator visitation and fruit set of avocado (*Persea americana* Mill.) in Kenya. *Arthropod-Plant Interact* 17:19–29. <https://doi.org/10.1007/s11829-022-09939-4>
- Tschumi M, Albrecht M, Bärtschi C et al (2016a) Perennial, species-rich wildflower strips enhance pest control and crop yield. *Agric Ecosyst Environ* 220:97–103. <https://doi.org/10.1016/j.agee.2016.01.001>
- Tschumi M, Albrecht M, Collatz J et al (2016b) Tailored flower strips promote natural enemy biodiversity and pest control in potato crops. *J Appl Ecol* 53:1169–1176. <https://doi.org/10.1111/1365-2664.12653>
- Tschumi M, Albrecht M, Entling MH, Jacot K (2015) High effectiveness of tailored flower strips in reducing pests and crop plant damage. *Proc R Soc B-Biol Sci* 282:20151369. <https://doi.org/10.1098/rspb.2015.1369>
- Tsuruda M, Girod P, Clausen M, Carrillo J (2023) Aromatic border plants in early season berries do not increase parasitism of spotted wing drosophila, *Drosophila suzukii*. *Pest Manage Sci* 79:134–139. <https://doi.org/10.1002/ps.7182>
- Tulli MC, Carmona DM, Vincini AM (2013) Influence of plant diversity on the numerical response of *Eriopsis connexa* (Coleoptera: Coccinellidae) to changes in cereal aphid density in wheat crops. *Int J Ecol* 2013:1–8. <https://doi.org/10.1155/2013/789532>
- Tyler-Julian K, Funderburk J, Srivastava M et al (2018) Evaluation of a push-pull system for the management of *Frankliniella* species (Thysanoptera: Thripidae) in tomato. *Insects* 9:187. <https://doi.org/10.3390/insects9040187>
- Udayakumar A, Shivalingaswamy TM, Bakthavatsalam N (2021) Legume-based intercropping for the management of fall armyworm, *Spodoptera frugiperda* L. in maize. *J Plant Dis Prot* 128:775–779. <https://doi.org/10.1007/s41348-020-00401-2>
- Uesugi R, Konishi-Furihata R, Tabuchi K et al (2023) Predacious natural enemies associated with suppression of onion thrips, *Thrips tabaci* (Thysanoptera: Thripidae), in intercropped onion-barley agroecosystems. *Environ Entomol* 52:183–196. <https://doi.org/10.1093/ee/nvad014>
- Varah A, Jones H, Smith J, Potts SG (2020) Temperate agroforestry systems provide greater pollination service than monoculture. *Agric Ecosyst Environ* 301:107031. <https://doi.org/10.1016/j.agee.2020.107031>
- Varchola JM, Dunn JP (2001) Influence of hedgerow and grassy field borders on ground beetle (Coleoptera: Carabidae) activity in fields of corn. *Agric Ecosyst Environ* 83:153–163. [https://doi.org/10.1016/S0167-8809\(00\)00249-8](https://doi.org/10.1016/S0167-8809(00)00249-8)
- Venturini EM, Drummond FA, Hoshida AK, et al (2017) Pollination Reservoirs in Lowbush Blueberry (Ericales: Ericaceae). *J Econ Entomol* tow285. <https://doi.org/10.1093/jee/tow285>
- Vidal S (1997) Factors influencing the population dynamics of *Brevicoryne brassicae* in undersown Brussels sprouts. *Biol Agric Hortic* 15:285–295. <https://doi.org/10.1080/01448765.1997.9755204>

- Vu Q, Ramal AF, Villegas JM et al (2018) Enhancing the parasitism of insect herbivores through diversification of habitat in Philippine rice fields. *Paddy Water Environ* 16:379–390. <https://doi.org/10.1007/s10333-018-0662-y>
- Wale M, Schulthess F, Kairu EW, Omwega CO (2007) Effect of cropping systems on cereal stemborers in the cool-wet and semi-arid ecozones of the Amhara region of Ethiopia. *Agric for Entomol* 9:73–84. <https://doi.org/10.1111/j.1461-9563.2007.00324.x>
- Wan N-F, Su H, Cavalieri A et al (2020b) Multispecies co-culture promotes ecological intensification of vegetable production. *J Cleaner Prod* 257:120851. <https://doi.org/10.1016/j.jclepro.2020.120851>
- Wan N-F, Zhang Y-M, Huang K-H et al (2016) Ecological engineering of trap cropping promotes biocontrol services in peach orchard ecosystems. *Ecol Eng* 90:427–430. <https://doi.org/10.1016/j.ecoleng.2016.01.045>
- Wang G, Cui L-L, Dong J et al (2011) Combining intercropping with semiochemical releases: optimization of alternative control of *Sitobion avenae* in wheat crops in China: effects of intercropping with MeSA on *Sitobion avenae*. *Entomol Exp Appl* 140:189–195. <https://doi.org/10.1111/j.1570-7458.2011.01150.x>
- Wang J, Li S, Fang Y et al (2022) Enhanced and sustainable control of *Myzus persicae* by repellent plants in organic pepper and eggplant greenhouses. *Pest Manage Sci* 78:428–437. <https://doi.org/10.1002/ps.6681>
- Wangai PW, Burkhard B, Müller F (2016) A review of studies on ecosystem services in Africa. *Int J Sustainable Built Environ* 5:225–245. <https://doi.org/10.1016/j.ijsbe.2016.08.005>
- Weißinger L, Schrieber K, Breuer M, Müller C (2019) Influences of blackberry margins on population dynamics of *Drosophila suzukii* and grape infestation in adjacent vineyards. *J Appl Entomol* 143:802–812. <https://doi.org/10.1111/jen.12669>
- Westich R, Hough-Goldstein J (2001) Temperature and host plant effects on predatory stink bugs for augmentative biological control. *Biol Control* 21:160–167. <https://doi.org/10.1006/bcon.2001.0928>
- Winkler K, Wäckers FL, Termorshuizen AJ, Van Lenteren JC (2010) Assessing risks and benefits of floral supplements in conservation biological control. *Biocontrol* 55:719–727. <https://doi.org/10.1007/s10526-010-9296-8>
- Woltz JM, Isaacs R, Landis DA (2012) Landscape structure and habitat management differentially influence insect natural enemies in an agricultural landscape. *Agric Ecosyst Environ* 152:40–49. <https://doi.org/10.1016/j.agee.2012.02.008>
- Woodcock BA, Bullock JM, McCracken M et al (2016) Spill-over of pest control and pollination services into arable crops. *Agric Ecosyst Environ* 231:15–23. <https://doi.org/10.1016/j.agee.2016.06.023>
- Wu K, Jiang C, Zhou S, Yang H (2022) Optimizing arrangement and density in maize and alfalfa intercropping and the reduced incidence of the invasive fall armyworm (*Spodoptera frugiperda*) in southern China. *Field Crops Res* 287:108637. <https://doi.org/10.1016/j.fcr.2022.108637>
- Wu Y, Cai Q, Lin C et al (2009) Responses of ground-dwelling spiders to four hedgerow species on sloped agricultural fields in Southwest China. *Prog Nat Sci* 19:337–346. <https://doi.org/10.1016/j.pnsc.2008.05.032>
- Xiang H, Lan N, Wang F et al (2021) Reduced pests, improved grain quality and greater total income: benefits of intercropping rice with *Pontederia cordata*. *J Sci Food Agric* 101:5907–5917. <https://doi.org/10.1002/jsfa.11243>
- Xie H-C, Chen J-L, Cheng D-F et al (2012) Impact of wheat-mung bean intercropping on English grain aphid (Hemiptera: Aphididae) populations and its natural enemy. *J Econ Entomol* 105:854–859. <https://doi.org/10.1603/EC11214>
- Xu QC, Xu HL, Qin FF et al (2010) Relay-intercropping into tomato decreases cabbage pest incidence. *J Food Agric Environ* 8:1037–1041
- Xue Z, Peng T, Liu B et al (2023) Licorice strips enhance predator-mediated biological control in China's cotton crop. *Pest Manage Sci* 79:781–791. <https://doi.org/10.1002/ps.7243>
- Yang Q, Li Z, Ouyang F et al (2023a) Flower strips promote natural enemies, provide efficient aphid biocontrol, and reduce insecticide requirement in cotton crops. *Entomol Gen* 43(2):421–432. <https://doi.org/10.1127/entomologia/2022/1545>
- Yang Y, Zhang Y, Zhang J et al (2023b) Plant volatiles mediate *Aphis gossypii* settling but not predator foraging in intercropped cotton. *Pest Manage Sci* 79:4481–4489. <https://doi.org/10.1002/ps.7650>
- Yao F-L, You M-S, Vasseur L et al (2012) Polycultural manipulation for better regulation of planthopper populations in irrigated rice-based ecosystems. *Crop Prot* 34:104–111. <https://doi.org/10.1016/j.cropro.2011.12.003>
- Zada H, Saljoqi A-R (2019) Habitat manipulation through intercropping for the management of codling moth *Cydia pomonella* (Linnaeus) (Lepidoptera: Tortricidae) in Swat Pakistan. *Pak J Zool* 51. <https://doi.org/10.17582/journal.pjz/2019.51.4.1537.1545>
- Zarei E, Fathi SAA, Hassanpour M, Golizadeh A (2019) Assessment of intercropping tomato and sainfoin for the control of *Tuta absoluta* (Meyrick). *Crop Prot* 120:125–133. <https://doi.org/10.1016/j.cropro.2019.02.024>
- Zhang R, Ren L, Wang C et al (2004) Cotton aphid predators on alfalfa and their impact on cotton aphid abundance. *Appl Entomol Zool* 39:235–241. <https://doi.org/10.1303/aez.2004.235>
- Zhang W, Zhang T-T, Machado RAR, Dai C-C (2024) Intercropping-induced leaf metabolic changes increase plant resistance to herbivory. *Plant Soil*. <https://doi.org/10.1007/s11104-023-06437-1>
- Zhang X, Lövei GL, Ferrante M et al (2020) The potential of trap and barrier cropping to decrease densities of the whitefly *Bemisia tabaci* MED on cotton in China. *Pest Manage Sci* 76:366–374. <https://doi.org/10.1002/ps.5524>
- Zhang X, Ouyang F, Su J et al (2022) Intercropping flowering plants facilitate conservation, movement and biocontrol performance of predators in insecticide-free apple orchard. *Agric Ecosyst Environ* 340:108157. <https://doi.org/10.1016/j.agee.2022.108157>
- Zhang Z, Zhou C, Xu Y et al (2017) Effects of intercropping tea with aromatic plants on population dynamics of arthropods in Chinese tea plantations. *J Pest Sci* 90:227–237. <https://doi.org/10.1007/s10340-016-0783-2>
- Zhao JZ, Ayers GS, Grafius EJ, Stehr FW (2017) Effects of neighboring nectar-producing plants on populations of pest Lepidoptera and their parasitoids in broccoli plantings. *The Great Lakes Entomologist* 25. <https://doi.org/10.22543/0090-0222.1791>
- Zheng Y, Zhang L, Chen B et al (2020) Potato/Maize intercropping reduces infestation of potato tuber moth, *Phthorimaea operculella* (Zeller) by the enhancement of natural enemies. *J Integr Agric* 19:394–405. [https://doi.org/10.1016/S2095-3119\(19\)62699-7](https://doi.org/10.1016/S2095-3119(19)62699-7)
- Zhou H, Chen J, Liu Y et al (2013a) Influence of garlic intercropping or active emitted volatiles in releasers on aphid and related beneficial in wheat fields in China. *J Integr Agric* 12:467–473. [https://doi.org/10.1016/S2095-3119\(13\)60247-6](https://doi.org/10.1016/S2095-3119(13)60247-6)
- Zhou H, Chen L, Chen J et al (2013b) Adaptation of wheat-pea intercropping pattern in China to reduce *Sitobion avenae* (Hemiptera: Aphididae) occurrence by promoting natural enemies. *Agroecol Sustain Food Syst* 37:1001–1016. <https://doi.org/10.1080/21683565.2013.763887>

- Zhou Z, Chen Z, Xu Z (2010) Potential of trap crops for integrated management of the tropical armyworm, *Spodoptera litura* in tobacco. *J Insect Sci* 10:1–11. <https://doi.org/10.1673/031.010.11701>
- Zongo JO, Vincent C, Stewart RK (1993) Effects of intercropping sorghum-cowpea on natural enemies of the sorghum shoot fly, *Atherigona soccata* (Diptera: Muscidae), in Burkina Faso. *Biol Agric Hortic* 9:201–213. <https://doi.org/10.1080/01448765.1993.9754636>
- Zou Y, Shen F, Zhong Y et al (2022) Impacts of intercropped maize ecological shading on tea foliar and functional components, insect pest diversity and soil microbes. *Plants* 11:1883. <https://doi.org/10.3390/plants11141883>
- Zuma M, Njekete C, Konan KAJ et al (2023) Companion plants and alternative prey improve biological control by *Orius laevigatus* on strawberry. *J Pest Sci* 96:711–721. <https://doi.org/10.1007/s10340-022-01570-9>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.